

# Potential Applications of Nanotechnology in Agriculture: Conceptions, Characteristics, Prospects, and Limitations - A Review

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## Abstract

Nanotechnology is a trending area of multidisciplinary research. It provides many opportunities in diverse field of science like chemical engineering, medicine, pharmaceuticals, environment, and agriculture. The prospective uses and advantages of nano science in agriculture are enormous. Nanomaterials have small size and distinctive physical and chemical features. Due to this, they play significant role in the different agricultural applications like seed germination, seedling growth, plant development, and plant protection. Various studies show that technology based on nanomaterials will have large and deep-rooted impact on agricultural area and crop production. Nanomaterials improve seed germination percentage rate and increase length of root and shoot with their ratio and biomass. Nanomaterials enhance biological parameters such as photosynthetic process and nitrogen metabolism in several crop plants. In the recent agricultural scenario, nanotechnology is playing a major role in crop disease management because of its environment friendly nature and potentiality. To reduce the side effects of nanotechnology on health of living organism and environment, the world is moving towards green nanotechnology. Green synthesis of metallic nanoparticles, characterization of synthesized nanoparticles, and their potential application were discussed in this review.

## Keywords

Agriculture, Environment, Nanomaterials, Nanotechnology

## Introduction

Agriculture plays a vital character in the economy of a nation [1]. The increase in agricultural production is important for the development of a country [2]. Gross Domestic Product growth of a nation depends on growing food production rates [3]. Several factors such as climate change, soil nutrients, temperature, moisture content of air, and water holding capacity of soil directly affect food production and agricultural growth [4]. Thus, it is essential for a country to control the adversarial factors of agriculture [5]. Government needs to provide sustainable agriculture management as constantly growing population is raising the demand for higher agricultural yields and better strategies for optimization of agricultural practice [6]. The main purpose of sustainable agriculture for society is to provide best facilities for agriculture output and textile production [7]. There has been increased in crop production, soil nutrient, nitrogen efficiency, and water absorption with the use of sustainable agriculture practices [8]. Sustainable agriculture has promoted judicious use of fertilizers, herbicides, pesticides [9], and agrochemicals in some livestock production practices [10]. Nanotechnology is a modern technology which has shown its abilities in numerous arenas like environment, solar, medicine, engineering, agriculture, and pharmaceuticals [11-13]. It has been drawing attention for the last two decades and is currently

being heavily used in agriculture science and medical science [14]. Nanotechnology can positively impact sustainable agriculture with maximizing crop yields with minimum use of fertilizers, pesticides, and herbicides by observing ecological variables and applying directed action [15]. It can boost the agricultural production and its quality in environmental-friendly manner by giving innovative solutions to remediate and protect ecosystem [16]. Agricultural nanotechnology is a part of agricultural science that uses nanomaterials to improve agronomic management of soil, water, crops, and food [17]. Nanomaterials may have specific surface components, various types and concentrations of sites, and varied reactivity with reference to mechanisms like adsorption and electrochemical reactions, as compared to bulk materials, which could be useful in producing nanomaterials for agricultural purpose [18]. The huge surface area provided by small nanomaterials makes them desirable for overcoming difficulties that are not resolved by other control approaches [19]. As a result, this technology promotes in the fall of environmental hazards [20]. Nanotechnology becomes popular due to its potential applications in various sectors such as agriculture, healthcare, and the ecosystem. [13]. Nanotechnology is a current scientific discipline that explores nanoscale materials [21]. Nanomaterials can be well-definite as materials which have, at least, one outer dimension of 1 - 100 nm [22]. These have many advantages in agricultural and environmental application due to their distinctive chemical, physical, optical, and magnetic composition [23] that are controlled by their unique size and shape [24].

Nanomaterials are of different shapes which can be classified on the basis of their dimensionality [25]: a) with three dimensions fall in the nanoscale range are called zero-dimensional nanomaterials [26] such as quantum dots, and nanoparticles [27], b) with one dimension in the nanoscale range are called one-dimensional nanomaterials such as nanowires, nanotubes, and nanorods [28], c) with two dimensions in the range of nanoscale are called two-dimensional nanomaterials such as bundle of nanowires, and bundle of nanotubes [29], and d) in three dimensional nanomaterials, none of the dimensions fall within the nanoscale range such as nanofilms and nanolayers [30].

Nanotechnology offers its own set of advantages and drawbacks [31]. Natural sources and industrial manufacturing both contribute to the occurrence of nanomaterials in the surroundings [32]. Nanomaterials contact the land through accumulation in the environment and rain, as well as straight delivery in agricultural areas. Plants absorb nanoparticles from the soil and nanoparticles can also enter organisms. The world is heading into green nanotechnology in order to lessen the adverse effects of nanotechnology on health of human [33] and the surroundings [34]. The combination of nanotechnology functions with green chemistry techniques become a key element of the nanotechnology future [35]. Biogenic synthesis of metallic nanoparticles includes various biogenic materials such as bacteria, fungi, algae, enzymes [36], extracts of plants, and agricultural wastes [37]. Natural products with various chemical substances perform as suitable reducing agents for the synthesis of metallic nanoparticles [38]. Green

nanotechnology is required to avoid the construction of undesirable and harmful by-products through the authentic, sustainable, and environmentally friendly methods of synthesis [39]. These methods are beneficial since they are cheap, simple, and take less reaction time [40]. Green synthesis of nanoparticles includes the use of standard solvent systems and natural sources which is necessary to complete this goal [41]. Among the existing green methods to produce metal and metal oxide nanoparticles, plant extracts [42] and agricultural waste [43] are considered as one of the most favorable natural reducing agents [44]. Use of plant extracts and agricultural waste is a very simple and easy method to produce nanomaterials at large scale as compared to micro-organism mediated synthesis [45]. Nanomaterials developed using environment ecofriendly techniques can improve agriculture by improving fertilization [46], biological control agents [47], pesticide transportation of bioactive constituents to desired target locations [48], sewage treatment, and nutrient absorption in plants [49]. Additionally, they reduce the number of toxic substances out into the environment [50].

## Biogenic Synthesis of Metallic Nanoparticles

Recent studies reported that top-down and bottom-up techniques are frequently used to synthesize metal and metal oxide nanoparticles [51]. The size of bulk structures is condensed to the nanometre scale in the "top-down" technique [52], whereas the molecular structure of metallic nanoparticles is formed by organizing atoms or molecules in the "bottom-up" approach [53]. For metallic nanoparticle synthesis, the top-down approach covers physical methods, whereas the bottom-up approach involves biological and chemical methods [54]. The application of high temperature, high pressure, and hazardous substances in physical and chemical methods of nanoparticle manufacturing is damaging to the environment [55]. It was reported that natural products can create a variety of distinct nanostructures [56]. Many bacteria, fungi, and plants have demonstrated the potential to synthesize metallic nanoparticles, each with their own set of benefits and drawbacks [57]. Bio reduction, also known as the green method for production of metallic nanoparticles, is a bottom-up strategy in which plant extracts or microorganisms are employed to reduce metal salts to nanometer-sized metals [58]. The nanoparticles produced with this technology are harmless and stable [59]. Nanostructures with desired shapes and composition can be produced by regulating biogenic synthesis [60]. This has attracted scientists' attention in using natural products to synthesize nanostructures for a variety of uses.

## Synthesis of Metallic Nanoparticles Using Microorganism

Recent studies have shown that numerous microorganisms can synthesize metallic nanoparticles and can be utilized and transformed to improve their performance for this purpose [61, 62] There are two types of methods for the biogenic synthesis of metallic nanoparticles from micro-organism

[63]. The first is bio reduction, which involves chemical reduction of metal ions into more stable structures biologically [64]. Many organisms are capable of metal reduction, which involves the reduction of a metal ion and the oxidation of an enzyme [65]. This method produces inert and stable metallic nanoparticles that can be securely separated from an impure sample [57]. Biosorption is the second method [66]. This method involves the attachment of metal ions from a liquid or soil sample to the organism itself, such as the cell membrane, and this method works without the need for energy [67]. The following sections will provide an overview on the synthesis of metallic nanoparticles by bacteria and fungi.

### Synthesis using bacteria

Bacterial species has been widely studied as a source of metallic nanoparticles because of their abundance in the environment [68] and their capacity to adapt to extreme environmental conditions [69]. They're also rapidly growing, low-cost to grow, and simple to regulate [70]. Temperature, humidity, and incubation time may be easily controlled in bacterial growth during biogenic synthesis of metallic nanoparticles [71]. Intracellular and extracellular mechanisms have been used by bacteria to assist in the production of nanoparticles. The extracellular synthesis method has the potential of being quicker than intracellular synthesis because it wouldn't require a subsequent procedure to collect nanoparticles from microorganisms [72]. Bacteria are favored for nanoparticle production due to the minimal conditions required, flexibility of purification, and large inventory [73]. As a result, bacteria seem to be the most studied microorganisms [74]. To produce silver (Ag) metallic nanoparticles, *Bacillus mojavensis* BTCB15 [75], *Bacillus subtilis* [76] and *Pseudomonas aeruginosa* [77] bacterial species were utilized. *Bacillusendo phyticus* has been employed

to make Ag metallic nanoparticles with a diameter varying between 5 and 5.5 nm in recent years [78]. *Aeromonas hydrophila* as well as *Lactobacillus* sp., on the other hand, produce copper sulphide (CuS) [65] and titanium dioxide (TiO<sub>2</sub>) nanoparticles [79]. Gold (Au) nanoparticles were synthesized by *Micrococcus yunnanensis* [70] and *Mycobacterium* sp. [80] in recent studies while cadmium nanoparticles were synthesized by *Clostridium thermoaceticum* and *Escherichia coli*. [81]. Bacterial species can be utilized as catalysts to produce inorganic nanoparticles as well as active supporters in the synthesis of nanoparticles [82]. Bacteria can produce extracellular or intracellular nanomaterials in nutrient broth during the gestation period. As a result of this process, bacterial biosynthesis of metallic nanoparticles is a feasible, efficient, and appropriate method for manufacturing [83]. The size, synthesis method, and various applications of metallic nanoparticles produced by different bacterial species were summarized in Table 1.

### Synthesis using fungi

Fungi have been used to produce metallic nanoparticles, and they are attracting a lot of attention since they have some advantages over bacteria for nanoparticle synthesis [84]. It has been reported that fungi are viable extracellular enzyme secretors, and many of their species grow quickly, making laboratory culturing and storage simple [85]. As a result, fungus can produce metal nanoparticles and nanostructures either intracellularly or extracellularly by using reducing enzymes [86]. One of several key benefits of extracellular production of metal and metal oxide nanoparticles by fungus is the appearance of a huge number of enzymes in a freed and purified condition that can be applied directly for downstream preparation [87]. The simplicity of scaling up and downstream processing, financial sustainability, and the presence of myce-

**Table 1:** Synthesis of metallic nanoparticles using bacteria.

Bacterial Species	Metallic nanoparticles	Size (nm)	Synthesis method	Application	References
<i>Pseudomonas aeruginosa</i>	Ag	50 - 85	Extracellular	Antibacterial activity	[77]
<i>Bacillus subtilis</i>	Ag	3 - 20	Extracellular	Antibacterial activity	[76]
<i>Pantoea ananatis</i>	Ag	8.06 – 91.31	Extracellular	Antimicrobial activity	[74]
<i>Bacillus endophyticus</i>	Ag	5.1	Extracellular	Antimicrobial activity	[78]
<i>Bacillus mojavensis</i> BTCB15	Ag	2.3	Extracellular	Antimicrobial activity	[75]
<i>Shewanella loibica</i> PV-4	Cu	10 - 16	Extracellular	Antibacterial activity	[63]
<i>Shewanella loibica</i> PV-4	Pd	4 - 10	Extracellular and intracellular	Catalytic Activity	[61]
<i>Shewanella loibica</i> PV-4	Pt	2 – 7	Extracellular	Catalytic Activity	[83]
<i>Micrococcus yunnanensis</i>	Au	53.8	Extracellular	Cytotoxic activity and antibacterial activity	[70]
<i>Mycobacterium</i> sp.	Au	5 – 55	Extracellular	Anticancer activity	[80]
<i>Bacillus megaterium</i> NCIM2326	ZnO	45 – 95	Extracellular	Antimicrobial activity	[69]
<i>Halomonas elongata</i> IBRC-M 10214	ZnO	18.11	Extracellular	Antibacterial activity	[73]
<i>Acinetobacter schindleri</i> SIZ7	ZnO	20 - 100	Extracellular	Antimicrobial activity	[64]
<i>Aeromonas hydrophila</i>	CuS	200	Extracellular and intracellular	Antibacterial, anti-inflammatory, and antioxidant activity	[65]
<i>Bacillus cereus</i> MN181367	ZnO	58.77 - 63.3	Extracellular	Antibacterial activity	[68]

lia, which gives a bigger surface area, were discovered to be all important concerns [88]. Fungi-derived nanoparticles could be employed in the delivery of drugs [89], treatment for cancer [90], MRI and medical image processing [91], and other applications. Fungi-based nanoparticle synthesis is an innovative, cost-effective, and environmentally acceptable method [92]. In the past few years, a range of different fungi have been used to make metallic nanoparticles [93]. For example, *Rhizopus stolonifera* [94], *Rhodotorula glutinis*, and *Arthroderma fulvum* [95] fungal species were used to make Ag metallic nanoparticles. In recent times, *Colletotrichum* sp. has been used to generate metallic nanoparticles of aluminium oxide (Al<sub>2</sub>O<sub>3</sub>) by an extracellular method with a diameter ranging from 30 to 50 nm [96]. In recent investigations, *Aspergillus niger* and *Candida albicans* [97] synthesised zinc oxide (ZnO) nanoparticles, while *Penicillium chrysogenum* [98] and *Pleurotus ostreatus* [99] synthesised magnesium oxide (MgO) and Au nanoparticles. The *Fusarium oxysporum* fungus is linked to the synthesis of Au metallic nanoparticles with an ideal size of about 20 nm and a sphere-shaped shape [100]. Kobashiwaga et al. published the biogenic production of Ag nanoparticles using the lignolytic fungus *Trametes troglia*. And after 20 days, adding Ag salt of nitrate to the fungal solution does not lead to the formation of Ag metallic nanoparticles. However, under an alkaline pH of 13, the production of Ag nanoparticles is more durable for 72 hours due to the electrostatic adsorption of hydroxides on Ag nanoparticles [101]. Purified organic extracts of *Aspergillus terreus* [102] and *Aspergillus niger* [103], isolated from a medicinal plant, could be used to build a sustainable and environmentally friendly fungus nanofactory [104,105]. *Trichoderma viridae* was utilized to prepare a quick and environmentally benign technique of synthesizing

metallic nanoparticles by generating secondary metabolites at high temperature [106,107]. The size, synthesis method, and various applications of metallic nanoparticles produced by different fungal species were summarized in Table 2.

### Synthesis of Metallic Nanoparticles Using Plant Extract

Over the last decade, there has been an increase in studies into the biogenic synthesis of metal [108] and metal oxide nanoparticles [109] utilizing plants or plant extracts. For nanoparticle synthesis, techniques such as thermal evaporation, reduction using mild chemicals [110], photochemical reagents [111], and electrochemical reagents has mostly been used [112]. Although all these techniques are capable of generating nanoparticles, they have a number of drawbacks [113], including the high cost of the process and the fact that they are not eco-friendly due to the use of hazardous solvents and reductants [114]. Even though chemical modifiers are consumed more than plant extracts, these substances are toxic to the environment and have health impacts [115]. Green chemistry techniques are used to synthesize nanoparticles that are easy, accessible, less energy-intensive [116], eco-friendly, minimize the use of hazardous components [117], and improve the efficiency of the process to avoid these problems [118]. Polyphenolic compounds obtained from natural sources seem to be the most enormous organic antioxidants [119], with huge potential as medicines, vitamins, and dietary supplements [120]. The polyphenols present in plant components act as biological reducing agents as well as nanoparticle fixers, which is the basic principle in green synthesis techniques [121]. Plant components, including flowers, leaves, roots, and stems, have been actively used for biogenic synthesis due to

**Table 2:** Synthesis of metallic nanoparticles using fungi.

Fungal species	Metallic Nanoparticles	Size (nm)	Synthesis Method	Application	References
<i>Rhizopus stolonifer</i>	Ag	5 - 50	Extracellular	Anticancer	[88]
<i>Trichoderma longibrachiatum</i>	Ag	10	Extracellular	Antimicrobial activity	[92]
<i>Aspergillus terreus</i>	Ag	16 - 57	Extracellular	Antibacterial activity	[102]
<i>Rhodotorula glutinis</i>	Ag	15.45	Extracellular	Antifungal activity, and cytotoxicity	[66]
<i>Arthroderma fulvum</i>	Ag	15.5	Extracellular	Antifungal activity	[95]
<i>Fusarium oxysporum</i>	Ag	5 - 13	Extracellular	Antibacterial and antitumor activity	[93]
<i>Trichoderma harzianum</i>	Ag	20 - 30	Extracellular	Antifungal activity	[106]
<i>Cladosporium cladosporioides</i>	Ag	100	Extracellular	Antibacterial and antioxidant activities	[86]
<i>Pleurotus ostreatus</i>	Au	10 - 30	Extracellular	Antimicrobial and anticancer	[90]
<i>Aspergillus niger</i>	Ag	10 - 100	Extracellular	Antifungal activity	[103]
<i>Penicillium chrysogenum</i>	MgO	7 - 40	Extracellular	Antimicrobial activity	[98]
<i>Aspergillus niger</i>	ZnO	80 - 130	Extracellular	Therapeutic application	[91]
<i>Candida albicans</i>	ZnO	25	Extracellular	Synthesis of pyrazolins	[97]
<i>Fusarium keratoplasticum strain</i>	ZnO	10 - 42	Extracellular	Antibacterial and antioxidant activities	[84]
<i>Colletotrichum</i> sp.	Al <sub>2</sub> O <sub>3</sub>	30 - 50	Extracellular	Antimicrobial activity	[96]



the high-quality phytochemicals they generate [122]. In the biopharmaceutical fields, metallic nanoparticles of Ag can be extracted from medicinal herbs, including *Saccharum of cinarum*, *Cinamomum camphora*, *Oryza sativa*, *Medicago sativa*, and *Magnolia Kobus* [123]. The first work reported on Au nanoparticle synthesis employed geranium leaf extract as a capping and reducing agent. The terpenes in leaf extract were responsible for the reduction of Au ions to Au nanoparticles. These nanoparticles were said to be created in different shapes, including spherical, triangular, and decahedral [124]. *Gardenia jasminoides*, *Anogeissus latifolia*, *Glycine max*, *Musa paradisiaca*, *Doiropyros kaki*, and other plant extracts have been used in the biogenic production of palladium (Pd) and platinum (Pt) metallic nanoparticles [125]. Metallic nanoparticles of Cu [126] are prepared by reducing Cu salt with different plant extracts, like flower extracts of *Aloe vera*. Metallic oxide nanoparticles of tin [127] and titanium [128] were manufactured with an environmentally sustainable approach from the roots of *Rheum emodi*, a Himalayan herb that has gained a lot of interest in the medical field. Goutam et al. constructed metallic nanoparticles of TiO<sub>2</sub> from *Jatropha curcas* leaf extract [129], which has been confirmed by UltraViolet-Visible (UV-Vis) spectrophotometer, fourier transform infrared (FTIR) spectroscopy, and X-ray diffraction (XRD). Other studies suggest that plant extracts, rather than microbes, can be used to rapidly synthesize highly stable nanoparticles. A current study describes an environmentally friendly approach for biogenic synthesis of ZnO nanoparticles utilizing *Cassia renigera* bark extract, as well as an assessment of its impact on four rice cultivars (PR 7, basmati 1509, HR 47, and sharbati) [130]. The lack of pathogenicity is a key benefit of employing plants instead of bacteria or fungus to produce metallic nanoparticles [131]. As a result, the plant extract could be used in an appropriate mechanism for reducing and stabilizing metallic nanoparticles

[132]. The size, morphology, and various applications of metallic nanoparticles produced by different plant extract were summarized in Table 3.

### Synthesis of Metallic Nanoparticles Using Agricultural Waste

Agricultural waste is produced in hundreds of millions of tons per year [133]. It was reported that unlike other industrial wastes, it has a relatively pure composition [134] and does not contain highly hazardous components [135], resins [136], or other additives [137], which limits its potential for reuse in the environment [138]. In recent studies, it has been shown that various biotechnological procedures [139] have been developed to use agro-wastes to increase nutritional quality through solid state fermentation [140], enzyme production, organic acid production [141], and as renewable resources for biogas production [142]. It has been reported that the use of agro-wastes as biomolecule sources in green nanotechnology has got a lot of attention, with various agro-wastes being recorded for their potential in nano biotechnology [143]. In general, extracts containing active biomolecules that catalyze the creation of nanoparticles can be produced from dried and ground agro-waste materials [144] using a simple hot water extraction process. In the green synthesis of metal and metal oxide nanoparticles, plant secondary metabolites operate as stabilizing and reducing agents and used to make nanoparticles of metal oxides such as Zn [145], Au [146], and Pd [147]. The synthesized metallic nanoparticles were characterized by a UV-Vis spectrophotometer. Distinctive method for the biogenic synthesis of Ag metallic nanoparticles via agricultural waste has been reported recently [148]. They used an aqua mushroom extract of *Pleurotus giganteus* in AgNO<sub>3</sub> solution. Metallic nanoparticles of Ag are generated in a controlled manner in 3 – 4 hours at room temperature. Synthesized

**Table 3:** Synthesis of metallic nanoparticles using plant extract.

Plant Extract	Metallic nanoparticle	Size (nm)	Morphology	Applications	References
<i>Morus alba</i> L.	Ag	80 - 150	Spherical	Antibacterial activity	[42]
<i>Beta vulgaris</i> L.	Ag	20 - 50	Spherical	Anticancer activity	[108]
<i>Piper longum</i>	Ag	15 - 40	Spherical	Catalytic and antibacterial activity	[110]
<i>Cocos nucifera</i>	Ag	22	Spherical	Antimicrobial activity	[52]
<i>Emblica officinalis</i>	Ag	15	Spherical	Antibacterial activity	[54]
<i>Abelmoschus esculentus</i>	Au	45 - 75	Spherical	Antifungal activity	[116]
<i>Eucommia ulmoides</i>	Au	16.4	Spherical	Catalytic activity	[59]
<i>Morinda citrifolia</i>	Au	12.17 – 38.26	Spherical	Anticancer activity	[115]
<i>Solanum nigrum</i>	Au	50	Spherical	Antibacterial activity	[113]
<i>Zingiber officinale</i>	Au	5 – 15	Spherical	Biomedical	[131]
<i>Aloe barbadensis</i>	CuO	15 – 30	Spherical	Nanomaterial	[126]
<i>Gloriosa superba</i>	CuO	5 - 10	Spherical	Antibacterial activity	[109]
<i>Punica granatum</i>	Fe <sub>3</sub> O <sub>4</sub>	12.6	Spherical	Water purification	[60]
<i>Cassia fistula</i>	ZnO	5 - 15	Spherical	Photocatalyst, and antioxidant	[119]
<i>Moringa oleifera</i>	ZnO	24	Spherical	Photocatalytic activity	[132]

nanoparticles were spherical in shape, with an average particle size of 20 nm. Fruit husk of *Nypa fruticans*, a harmless, environmentally safe biological resource, was used to successfully produce metallic nanoparticles of Au. The synthesis process rate is relatively faster than that of other natural products. The synthesized Au nanoparticles were characterised by transmission electron microscopy (TEM), UV-Vis spectrophotometer, and XRD. In a study, rice bran was used as a precursor in the production of ZnO nanoparticles [149]. Rice bran is a cost-effective and biological precursor for Zn nanoparticles production among the suitable agricultural wastes. TEM was used to analyze the structure and morphology of the produced metallic nanoparticles. The production of metallic Ag nanoparticles was investigated by utilizing *Cocos nucifera* mesocarp layer extract [150]. When AgNO<sub>3</sub> solution was reacted with a dilute extract of *C. nucifera* coir at 60 °C, Ag ions were reduced. The activation of surface plasmon resonance using UV-Vis spectrophotometer [151] at 433 nm was used to confirm the synthesized Ag metallic nanoparticles. Sinsinwar et al. suggested that extracts from agricultural wastes were utilized as reducing and stabilizing agents for their syntheses in the majority of cases [152], with a wide range of actions ranging from antibacterial, antifungal, insecticidal, catalytic, and cytotoxicity against cancer cells. Because of its biomedical applications, nontoxicity, ecological approach, and green chemistry nature, phyto-synthesis of transition metal nanoparticles is getting more popular. The size, morphology, and various applications of metallic nanoparticles synthesized from agricultural waste were summarized in Table 4.

## Characterization of Metallic Nanoparticles

Nanomaterials have indeed been explored using such several techniques to detect their dimensions, morphology, crystallographic, chemical compositions, and other physical characteristics [153]. In many areas, physical aspects can indeed be assessed using more than one technique [154]. Size and shape are two of the most important criteria addressed in the characterization of nanoparticles [155]. Researchers also could analyze the surface morphology and estimate the size distribution, surface charge density, quantity of accumulation, and surface area [156]. Various approaches for characterization are classified focused on the technology's concept, the insights it may offer, and the components it is suitable for [157]. These characterization techniques are sometimes used specifically to investigate a specific parameter, while other times they are combined [158]. The thickness, concentration, and ligands present on the interfaces of nanostructures may affect their additional areas of characterization [159]. There are two types of major techniques that are widely used for the characterization of nanomaterials. The first is spectroscopic and the other one is microscopic.

### Spectroscopic techniques

Many spectroscopic techniques are used for the characterization of metallic nanoparticles, out of which XRD is most used technique [160]. XRD normally gives information about the structure, nature, parameters, and size of the crystal [161]. In XRD measurement, the crystal parameter is calculated through using Scherrer equation, which is focused on

**Table 4:** Synthesis of metallic nanoparticles using agricultural waste.

Agricultural waste	Metallic nanoparticle	Size (nm)	Morphology	Application	References
Watermelon rind	Pd	96	Spherical	Catalytic activity	[147]
<i>Lansium domesticum</i> fruit peel	Au	20 - 40	Triangular and hexagonal	Biomedicine	[124]
Acacia gum	MgO	<100	Spherical	Divalent metallic species removal from water	[136]
Grape waste	Au	20 - 25	Spherical	Biological applications	[137]
Banana peel extract	Ag	23.7	-	Antibacterial activity	[141]
Mushroom Extract of <i>Pleurotus giganteus</i>	Ag	2-20	Spherical	Therapeutic applications	[148]
Waste coconut husk	ZnO	10 - 100	Spherical	Photocatalytic activity	[138]
Orange peels	ZnO	30 - 100	Spherical	Antibacterial activity, antioxidant activity, and anticancer activity	[145]
Mandarin waste peels	Ag	10 - 19	Spherical/ Oval	-	[140]
Rice bran	ZnO	17.16	Spherical	Antibacterial activity	[149]
Hybrid grape pulp extract	ZnO	23.56	Irregular hexagonal	Photocatalytic degradation activity	[135]
Corn cob	Au	35	Spherical	Antibacterial activity	[139]
<i>Passiflora edulis</i> peels	Au	7	Spherical	Antibacterial effect and catalytic activity	[134]
<i>Nypa fruticans</i> fruit husk	Au	15 - 20	Spherical	Antibacterial activity	[146]
Pomegranate ( <i>Punica granatum</i> ) Extracts	ZnO	32.98	Spherical and hexagonal-shaped	Antibacterial Activity	[133]

the broadening of a sample's peak intensity [162]. Using X-ray line broadening, Holder et al. calculated the standard size of the crystal of metallic nanoparticles, which was found to be between 9 and 53 nm [163]. There are several different techniques that may be used to determine the basic characteristics of metallic nanoparticles like size, shape, structure, and composition. Another major analytical approach for determining the molecular structural properties of nanomaterials is nuclear magnetic resonance spectroscopy [164]. In recent studies, it has been reported that it is based on the phenomenon that occurs when spin active nuclei are positioned in a high external magnetic field, resulting in a slight energy divergence within the down and up spin states [165]. Xiang et al. published a detailed article about NMR techniques in novel metallic nanoparticles. It was explained that characterization of nanoscale materials is also done using the BET approach, named after the developers' surnames initials, Brunauer, Emmett, and Teller, and founded on the principle of the adsorption process of a gaseous phase on a solid surface [166]. It's used to find out the surface area of nanomaterials, and it's a rather precise, quick, and simple method for doing so [167]. UV-Vis spectrophotometer is yet another popular characterization approach for nanostructures that is both simple and inexpensive [164]. It measures the light output emitted by a substance in relation to the amount of light emitted by a reference material. R Begum et al. inferred that UV-Vis spectrophotometer is a useful method for identifying, investigating, and evaluating the stability of nanoparticle colloidal solutions because the optical properties of nanoparticles are affected by their dimension, morphology, quantity, and refractive index [168].

### Microscopic techniques

An important technique that is used to characterize the construction of different nanocomposites that can be similar in structure to numerous crystal systems is TEM [164]. It was reported that TEM is an important microscopy technology that can utilize advantage of the interaction between an electron beam with a constant current density and a sample. The amount of the interaction is determined by several factors, including sample size, density, and composition of element. The information obtained from the transmitted electrons is used to create the final image. An important technique for analyzing the morphology and structure of nanomaterials is the scanning electron microscope (SEM). In this technique, a low-energy beam of electrons is projected onto the substance and analyses the substance's surface. As the electron beam penetrates and hits the material, a number of interactions occur, resulting in the emission of electrons and protons from the surface of the sample. Sundar et al. employed a high-resolution SEM to visualize Au nanoparticles in tissues and cells [169]. This approach ensures the simple viewing of metallic nanoparticles, and preparation of sample is quick and painless. According to Gemmi et al., electron diffraction also known as selected area electron diffraction, is another fundamental microscopy technique for investigating the crystalline structure of nanoparticles [170]. Various parameters of metallic nanoparticles and their characterization techniques were summarized in Table 5.

## Role of Nanotechnology in Agriculture

Due to the restricted resources of petroleum fuels and natural gas, the cost of raw materials such as synthetic fertilizers and pesticides is predicted to rise at an alarming rate [3]. A precision agricultural method is a more suitable substitution to lessen the cost of crop production and improve output [4]. Because of advancements in nanotechnology,

**Table 5:** Characterization techniques for different parameters.

Parameters	Characterization techniques	References
Detection of nanoparticles	TEM, SEM, STEM, and magnetic susceptibility	[160]
Size and shape of nanoparticles	TEM, XRD, HRTEM, AFM, EPLS, FMR, and 3D-tomography	[157]
Crystal structure	XRD, HRTEM, electron diffraction, and STEM	[166]
Surface area and charge	BET, liquid NMR, Z-potential, and EPM	[162]
3D visualization	3D-tomography, AFM, and SEM	[164]
Optical character	UV-Vis spectrophotometer	[154]

a variety of methods for remodeling precision agricultural practices are now available, allowing effective control at the nanoscale [171]. In nanotechnology, agrochemical exhibits amazing qualities at the nanoscale that bulk materials do not [6]. Nanoscience can also improve the genetic code of plants by transporting chemical molecules and proteins to exact locations at the cell surface [31]. Excessive and improper use of pesticides and fertilizers in conventional methods has increased chemicals and pollutants in surface and groundwater [7], resulting in increasing healthcare and water purification expenditures as well as a reduction in fisheries and recreational areas [21]. The disadvantages of conventional technologies can be seen in the fact that proponents of alternative farming, such as "agricultural production", propose environments that are neither unique nor effective because farming operates in an open process, making conservation agriculture thermochemical unviable. With the use of nanomaterials, traditional fertilizers have been replaced by nanoscale fertilizers [14]. Nano-fertilizers tend to enhance soil fertility and assist in the reduction of anaerobic decomposition and environmental contamination [15]. Nowadays, the research technology development process seems to have the capability to increase agricultural efficiency, productivity, and quality of products and hence nutritive value [20]. In developed countries, nano-based biosensors and agrochemicals for agricultural applications are all being used. Nano-sensors, which could diagnose organic pollutants and pathogens, and nano-delivery processes that would accurately release the drug at the right time, are among the applications expected for food and agriculture [172].

## Applications of Nanotechnology in Sustainable Agriculture

Nanotechnology has recently been proven to have the ability to revolutionize the agriculture sector by increasing the efficiency of agriculture products [10] and providing solutions to environmental and agricultural challenges to improve food production and sustainability, according to recent studies [2].



To fulfill the demands of a growing population, nanoscience uncovers a broad range of unique functions in the sector of food and agriculture [8]. Nanotechnology has a wide range of applications, including the use of agricultural resources such as nutrients, chemicals, and water in agricultural processes [5]. Nanoparticles have also proven to be effective in the field of crop biotechnology, where they have been used to protect agricultural crops [1]. Different uses of nanomaterials in agriculture are thoroughly evaluated in this review, including nano-fertilizers, nano-pesticides, nano-biosensors, and nanomaterials for soil remediation via biotic and abiotic pathways (Figure 1).

## Nano-biosensors

Nano-biosensors serve as a steppingstone for basic research and provide instruments for authentic bio-analytical

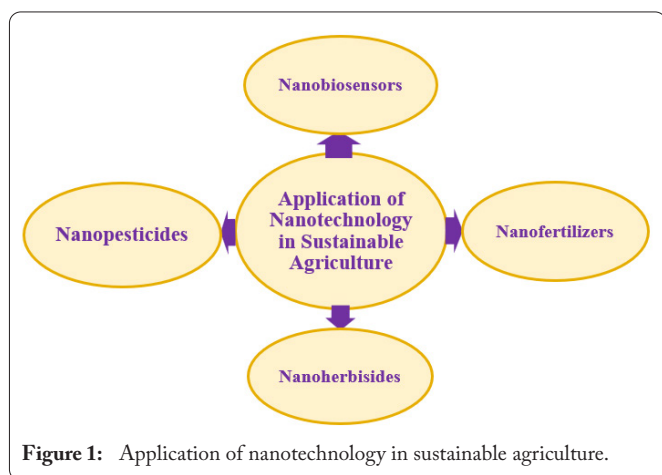


Figure 1: Application of nanotechnology in sustainable agriculture.

applications that were previously unattainable [4]. Nano-biosensors are nano-sensors immobilized with bioreceptors that are unique to the chemicals of interest [173]. A nano-biosensor is a device that uses nanotechnology to detect and evaluate data at the atomic level [6]. Biologically activated elements, a transducer, and a detector are the three main components of a nano-biosensor [174]. Neurotransmitters, proteins, antibodies, nucleotides, chemical impressions, and tissue that receive signals from the sample components and send them to the transducer are examples of biologically sensitive elements [47]. The transducer serves as an intermediary, monitoring the physical change caused by the reaction at the bioreceptor and converting that energy into a detectable electrical output [7]. The detector component captures the transducer's signals, which are then magnified and evaluated by a microcontroller before being transferred to a computer [175]. Nano-biosensors are currently fulfilling their potential application in agriculture. These provide accurate and timely information about the crop and its field condition with crop growth [176]. Excessive use of pesticides and fertilizers can also be reduced by real time monitoring which is useful in the decrease of ecological defect and manufacturing cost [11]. Nano-biosensors are also used to manage soil nutrients and to reduce pollution in the environment [177]. Nano-biosensors have various advantages, including increased sensitivity, which reduces the reaction time for detecting plant pathogens in crops, hence speeding agricultural productivity and minimizing food safety concerns [15].

These are useful in the detection of the humidity of soil, residue of pesticide, and concentration of nutrients [178]. These sensors are extremely sensitive, enable to detect single virus particles or extremely low amounts of a potentially dangerous material [179]. The traditional methods of agriculture are converting into smart methods using nano-sensors which are environment friendly and more efficient in term of energy for the sustainable agriculture [180]. Various types of nanomaterials can be used to make nano-biosensors such as quantum dots carbon nanotube Au nanoparticles and different nanocomposites with their polymer [181]. These are highly sensitive due to their large surface area. These showed quick response in seconds which is reliable and more stable [182]. Nano-biosensors have a lot of potential above regular biosensors right now [17]. These have distinct advantages, such as increased detection sensitivity, and have enormous potential for use in a variety of fields, including environmental and biological process regulation, food quality control, agriculture, biosafety, and most notably therapeutic applications.

## Nano-fertilizers

At present, conventional chemical fertilizers are being used by different methods to increase the productivity of the crop [18]. However, less than half of the chemical fertilizer used by the crop and the remaining quantity fixed inside the soil layers that cause soil pollution [183]. Chemical fertilizers are used repeatedly, due to which the fertility of the soil decreases as well as it loses the soil nutrient which are necessary for growing any crop [46]. Nanotechnology can adapt fertilizer manufacturing to the desired chemical composition, increase nutrient use efficiency while reducing environmental impact, and increase plant output [184]. A nano-fertilizer is a nutrient-delivery device that works in one of three different ways [185]. The nutrients could be enclosed inside nanomaterials such as nanoporous materials or nanotubes, wrapped in a thin polymeric membrane or supplied as nano nutrients [186]. In nano-fertilizers, nutrients are bonded to nano-sized adsorbents that distribute nutrients more gradually than conventional fertilizers, whether applied alone or in combination [187]. This method not only improves fertilizer utilization, but it also reduces nutrient loss into groundwater [188]. It can also be utilized to improve abiotic stress tolerance, and when combined with microorganisms, they provide even greater benefits [189]. Nano-fertilizers are made up of a variety of nanomaterials such as metals, oxides of metals, C-based compounds, and any other nanostructured elements, with various compositions and characteristics [190]. It can be made in a variety of ways, including top-down-a physical method, bottom-up-a chemical method, and other biological methods [19]. After years of scientific research based on particular nutrients, it has always been possible to develop different types of nano-fertilizers, such as micro-nutrient nano-fertilizers, macro-nutrient nano-fertilizers, and nanomaterial-enhanced fertilizers [191]. To reduce nutrient losses, present nano-sized formulations of nano-fertilizers improve solubility, diffusion of insoluble elements, bioavailability, and transparency of definite distribution [192]. Studies suggested that different nanoparticles such as Fe, Cu, Fe<sub>2</sub>O<sub>3</sub>, TiO<sub>2</sub>, ZnO, and CeO<sub>2</sub> can be used as nano-fertilizers [193]. At present, nano-fertilizers are being seen as a



promising alternative in agriculture as well as being considered better in every respect than conventional fertilizers [13]. Thus, based on the facts above, we can conclude that the use of nano-fertilizers increases the effectiveness of micro and macro elements, lowers soil toxicity, and reduces the frequency with which traditional fertilizers are applied.

### Nano-pesticides

Nano-pesticides are two to three-dimensional nanostructures with dimensions ranging from 1 to 200 nm that are used to transport agrochemical chemicals [48]. When compared to basic pesticides, putting agrochemical into nanoparticles offers advantages due to their specific features [32]. These pesticides formulated in nanomaterials for agricultural uses, whether particularly fixed on a hybrid base, enclosed in a matrix, or complexed nanocarriers for external stimulation or enzyme-mediated reactions [194]. Nanosized particles, in combination with their shape and specific features, are expected to investigate pesticide actions in nanocarrier novel formulations based on a variety of materials, including silica, fats, polysaccharides, co-polymers, porcelain, metals, charcoal, and others [195]. Nanopesticide compositions can improve solubility in water, absorption, and preserve agrochemicals from degradation in the environment, reinventing disease, weed, and insect control in crops [34]. The neurotoxicity and genotoxicity of nanomaterials, on the other hand, are on the edge [196]. Use of traditional pesticides that is uncontrolled and illogical disrupts the ecosystem's balance and puts everyone's life in danger [197]. An acute and chronic adverse effect of industrial or unintentional absorption of pesticide residues from food, water consumption is lethal [198]. Adolescents are more sensitive to pesticides and are more likely to suffer irreversible tissue and organ damage as a result [199]. The cerebral and peripheral neurotoxic effects, as well as the effect on blood coagulation capacity, are all valid reasons to be concerned [200]. Nano-pesticides from industrial and agricultural sewage runoff enter the water supply by soil penetrating seeping after a precipitation event, changing its quality, enhancing human exposure time, and raising issues for the ecosystem [201]. Nanotechnologies strive to limit the uncontrolled and harmful use of traditional pesticides while also ensuring their safe application [20]. The most significant technological development for the safe use of pesticides and innovative ideas to supply nano-pesticides novel material is grafted intended nanoparticle compositions for ecological stimulations.

### Nano-herbicides

Weeds are a major hazard in agriculture, as they drastically impair productivity. As a result, there is no choice except to eliminate them [9]. Nanotechnology can eliminate weeds in an environmentally acceptable manner by using nano-herbicides that do not leave hazardous residues in the soil or environment [12]. If the active component is paired with a "smart" delivery method, less herbicide will be utilized [35]. Because they are nanoscale in size, they will blend in with soil particles and hinder the growth of weed species that have developed resistance to traditional herbicides [23]. Herbicides on the market are designed to control or kill the weed plants above-ground parts [33]. Generating a specific herbicide component coated with nanoparticle that enters the root system of target weeds

and translocate to sections that impede glycolysis of food stores in the root system, causing the specific weed plant to go starved and die [37]. Since they are target-specific, nanoparticles may indeed be able to destroy weeds and enhance yields. There are also restrictions in place for the regulated release of herbicides, which limit human damage [36]. Herbicides such as ametryn, triazine, and atrazin have all been nano-encapsulated to achieve an effective plant absorption capacity of 84% [188]. Atrazine is a pesticide that is commonly used to destroy weeds and unwanted grass that grows near farms [202]. The adsorptive stripping voltammetry process was developed to synthesise the herbicide fenclorim using carbon nanotubes at pH 4 and adsorption tactics on the electrodes [203]. Herbicide use depletes soil minerals over time and renders crops resistant to herbicides. Nevertheless, utilizing modified silver with nanoparticles and carboxymethyl cellulose facilitates herbicide breakdown [41].

## Conclusions

In agriculture, nanotechnology has a variety of applications such as nano-fertilizers, nano-biosensors, nano-pesticides, etc. These are effective tools for smart agriculture systems because of their nanoscale-controlled release and site-specific delivery which overcome the side effects of conventional methods. Nano-sensors have demonstrated the ability to detect leftover pesticides, ensuring consumer food safety which is an advantage over to traditional methods. The application of metallic nanoparticles may improve the yield and development of crop plants, but the outcomes vary according to the species of plants. To understand the outcome of nanomaterials in the ecosystem, a long-term assessment of each crop-nanoparticles system is required. Within concentration levels, certain metal and metal oxide nanoparticles improved seed germination percentage, shoot and root elongation, biomass production, and plant development. Metallic nanoparticle screening and modification for numerous species of plants is needed for the commercialization of nanotechnology. The efficiency and utility of nanoparticles can indeed be controlled by modifying their compositions and durability. Over to conventional methods more research is needed to determine the expected level for each crop system.

## References

1. Bhati M. 2021. Biogenic synthesis of metallic nanoparticles: Principles and applications. *Materials Today: Proceedings*. <https://doi.org/10.1016/j.matpr.2021.04.272>
2. Zhao L, Lu L, Wang A, Zhang H, Huang M, et al. 2020. Nano-biotechnology in agriculture: use of nanomaterials to promote plant growth and stress tolerance. *J Agric Food Chem* 68(7): 1935-1947. <https://doi.org/10.1021/acs.jafc.9b06615>
3. Majumdar S, Keller AA. 2020. Omics to address the opportunities and challenges of nanotechnology in agriculture. *Critical Reviews in Environmental Science and Technology* 51(22): 2595-2636. <https://doi.org/10.1080/10643389.2020.1785264>
4. Prasad R, Bhattacharyya A, Nguyen QD. 2017. Nanotechnology in sustainable agriculture: recent developments, challenges, and perspectives. *Front Microbiol* 8: 1014. <https://doi.org/10.3389/fmicb.2017.01014>
5. Rane AV, Kanny K, Abitha VK, Thomas S. 2018. Methods for synthesis of nanoparticles and fabrication of nanocomposites. In: Bhagyaraj SM, Oluwafemi OS, Kalarikkal N, Thomas S (eds) *In micro and nano technologies, synthesis of inorganic nanomaterials*. Woodhead Publishing,

- Sawston, pp 121–139. <https://doi.org/10.1016/B978-0-08-101975-7.00005-1>
6. Usman M, Farooq M, Wakeel A, Nawaz A, Cheema SA, et al. 2020. Nanotechnology in agriculture: Current status, challenges and future opportunities. *Science of the Total Environment* 721: 137778. <https://doi.org/10.1016/j.scitotenv.2020.137778>
  7. Parisi C, Vignani M, Rodríguez-Cerezo E. 2015. Agricultural nanotechnologies: what are the current possibilities? *Nano Today* 10(2): 124–127. <https://doi.org/10.1016/j.nantod.2014.09.009>
  8. Chaudhry N, Dwivedi S, Chaudhry V, Singh A, Saquib Q, et al. 2018. Bio-inspired nanomaterials in agriculture and food: Current status, foreseen applications and challenges. *Microb Pathog* 123: 196–200. <https://doi.org/10.1016/j.micpath.2018.07.013>
  9. Pirzadah TB, Malik B, Maqbool T, Rehman RU. 2019. Development of nano-bioformulations of nutrients for sustainable agriculture. In: Prasad R, Kumar V, Kumar M, Choudhary D (eds) *Nanobiotechnology in Bioformulations. Nanotechnology in the Life Sciences*. Springer, Cham, pp 381–394. [https://doi.org/10.1007/978-3-030-17061-5\\_16](https://doi.org/10.1007/978-3-030-17061-5_16)
  10. Fu L, Wang Z, Dhankher OP, Xing B. 2020. Nanotechnology as a new sustainable approach for controlling crop diseases and increasing agricultural production. *J Exp Bot* 71(2): 507–519. <https://doi.org/10.1093/jxb/erz314>
  11. Iavicoli I, Leso V, Beezhold DH, Shvedova AA. 2017. Nanotechnology in agriculture: Opportunities, toxicological implications, and occupational risks. *Toxicol Appl Pharmacol* 329: 96–111. <https://doi.org/10.1016/j.taap.2017.05.025>
  12. Pudake RN, Chauhan N, Kole C, editors. 2019. *Nanoscience for sustainable agriculture*. Springer International Publishing, Switzerland.
  13. Martins R, Kaczerewska OB. 2021. Green nanotechnology: The latest innovations, knowledge gaps, and future perspectives. *Appl Sci* 11(10): 4513. <https://doi.org/10.3390/app11104513>
  14. Ali S, Shafique O, Mahmood T, Hanif MA, Ahmed I, et al. 2018. A review about perspectives of nanotechnology in agriculture. *Pakistan Journal of Agricultural Research* 30(2): 116–121. <http://doi.org/10.17582/journal.pjar/2018/31.2.116.121>
  15. Worrall EA, Hamid A, Mody KT, Mitter N, Pappu HR. 2018. Nanotechnology for plant disease management. *Agronomy* 8(12): 285. <https://doi.org/10.3390/agronomy8120285>
  16. Bueno PD, Gillerman L, Gehr R, Oron G. 2017. Nanotechnology for sustainable wastewater treatment and use for agricultural production: A comparative long-term study. *Water Research* 110: 66–73. <https://doi.org/10.1016/j.watres.2016.11.060>
  17. Marchiol L, Iafisco M, Fellet G, Adamiano A. 2020. Nanotechnology support the next agricultural revolution: Perspectives to enhancement of nutrient use efficiency. In: Sparks DL (ed) *Advances in Agronomy*. Academic Press, pp 27–116. <https://doi.org/10.1016/bs.agron.2019.12.001>
  18. Marslin G, Siram K, Maqbool Q, Selvakesavan RK, Kruszka D, et al. 2018. Secondary metabolites in the green synthesis of metallic nanoparticles. *Materials (Basel)* 11(6): 940. <https://doi.org/10.3390/ma11060940>
  19. Chandra H, Kumari P, Bontempi E, Yadav S. 2020. Medicinal plants: Treasure trove for green synthesis of metallic nanoparticles and their biomedical applications. *Biocatalysis and Agricultural Biotechnology* 24: 101518. <https://doi.org/10.1016/j.cbac.2020.101518>
  20. Pérez-de-Luque A. 2017. Interaction of nanomaterials with plants: what do we need for real applications in agriculture? *Front Environ Sci* 5: 12. <https://doi.org/10.3389/fenvs.2017.00012>
  21. Fraceto LF, Grillo R, de Medeiros GA, Scognamiglio V, Rea G, et al. 2016. Nanotechnology in agriculture: Which innovation potential does it have? *Front Environ Sci* 4: 20. <https://doi.org/10.3389/fenvs.2016.00020>
  22. Sudha PN, Sangeetha K, Vijayalakshmi K, Barhoum A. 2018. Nanomaterials history, classification, unique properties, production and market. In: Barhoum A, Hamdy Makhoul AS (eds) *Emerging applications of nanoparticles and architecture nanostructures*. Elsevier, pp 341–384. <https://doi.org/10.1016/B978-0-323-51254-1.00012-9>
  23. Kremer RJ. 2019. Bioherbicides and nanotechnology: current status and future trends. In: Koul O (ed) *Nano-biopesticides today and future perspectives*. Academic Press, pp 353–366. <https://doi.org/10.1016/B978-0-12-815829-6.00015-2>
  24. Saleh TA. 2020. Nanomaterials: Classification, properties, and environmental toxicities. *Environmental Technology & Innovation* 20: 101067. <https://doi.org/10.1016/j.eti.2020.101067>
  25. Singh V, Yadav P, Mishra V. 2020. Recent advances on classification, properties, synthesis, and characterization of nanomaterials. In: Srivastava N, Srivastava M, Mishra PK, Gupta VK (eds) *Green synthesis of nanomaterials for bioenergy applications*. Wiley, pp 83–97. <https://doi.org/10.1002/9781119576785.ch3>
  26. Dolez PI. 2015. Nanomaterials definitions, classifications, and applications. *Nanoengineering* pp 3–40. <https://doi.org/10.1016/B978-0-444-62747-6.00001-4>
  27. Wang Z, Hu T, Liang R, Wei M. 2020. Application of zero-dimensional nanomaterials in biosensing. *Front Chem* 8: 320. <https://doi.org/10.3389/fchem.2020.00320>
  28. Quan W, Xudong W, Min X, Lou X, Fan X. 2019. One-dimensional and two-dimensional nanomaterials for the detection of multiple biomolecules. *Chinese Chemical Letters* 30(9): 1557–1564. <https://doi.org/10.1016/j.ccl.2019.06.025>
  29. Tan C, Cao X, Wu XJ, He Q, Yang J, et al. 2017. Recent advances in ultrathin two-dimensional nanomaterials. *Chem Rev* 117(9): 6225–6331. <https://doi.org/10.1021/acs.chemrev.6b00558>
  30. Thirupathi AR, Sidhureddy B, Boateng E, Soldatov DV, Chen A. 2020. Synthesis and electrochemical study of three-dimensional graphene-based nanomaterials for energy applications. *Nanomaterials* 10(7): 1295. <https://doi.org/10.3390/nano10071295>
  31. Ditta A, Arshad M. 2016. Applications and perspectives of using nanomaterials for sustainable plant nutrition. *Nanotechnology Reviews* 5(2): 209–229. <https://doi.org/10.1515/ntrev-2015-0060>
  32. Chhipa, H. 2019. Applications of nanotechnology in agriculture. In: Gurtler G, Ball AS, Soni S (eds) *Methods in microbiology*. Academic Press, pp 115–142. <https://doi.org/10.1016/bs.mim.2019.01.002>
  33. Hamad HT, Al-Sharify ZT, Al-Najjar SZ, Gadooa ZA. 2020. A review on nanotechnology and its applications on Fluid Flow in agriculture and water recourses. *IOP Conf Ser: Mater Sci Eng* 870: 012038. <https://doi.org/10.1088/1757-899X/870/1/012038>
  34. Ahmad S, Munir S, Zeb N, Ullah A, Khan B, et al. 2019. Green nanotechnology: a review on green synthesis of silver nanoparticles - an ecofriendly approach. *Int J Nanomedicine* 14: 5087–5107. <https://doi.org/10.2147/ijn.s200254>
  35. Nasrollahzadeh M, Sajjadi M, Sajadi SM, Issaabadi Z. 2019. Green nanotechnology. In: *Interface science and technology*. Elsevier pp. 145–198. <https://doi.org/10.1016/B978-0-12-813586-0.00005-5>
  36. Pandey G. 2018. Challenges and future prospects of agri-nanotechnology for sustainable agriculture in India. *Environmental Technology & Innovation* 11: 299–307. <https://doi.org/10.1016/j.eti.2018.06.012>
  37. Yata VK, Tiwari BC, Ahmad I. 2018. Nanoscience in food and agriculture: research, industries and patents. *Environ Chem Lett* 16(1): 79–84. <https://doi.org/10.1007/s10311-017-0666-7>
  38. Guo C, Yarger JL. 2018. Characterizing gold nanoparticles by NMR spectroscopy. *Magn Reson Chem* 56(11): 1074–1082. <https://doi.org/10.1002/mrc.4753>
  39. Kakkar S, Harjani B, Ledwani N, Ledwani L. 2020. Synthesis, characterization, and application of biogenic nanomaterials: An overview. In: Ledwani L, Sangwai J (eds) *Nanotechnology for Energy and Environmental Engineering*, Springer, Cham pp 51–71. [https://doi.org/10.1007/978-3-030-33774-2\\_2](https://doi.org/10.1007/978-3-030-33774-2_2)
  40. Khadeeja P, Viktoria B, Lalita L. 2015. Synthesis of nanoparticles: their

- advantages and disadvantages. 2<sup>nd</sup> International Conference on Emerging Technologies: Micro to Nano 2015 (ETMN-2015).
41. Kanwar R, Rathee J, Salunke DB, Mehta SK. 2019. Green nanotechnology-driven drug delivery assemblies. *ACS Omega* 4(5): 8804-8815. <https://doi.org/10.1021/acsomega.9b00304>
  42. Razavi R, Molaei R, Moradi M, Tajik H, Ezati P, et al. 2020. Biosynthesis of metallic nanoparticles using mulberry fruit (*Morus alba* L.) extract for the preparation of antimicrobial nanocellulose film. *Applied Nanoscience* 10(2): 465-476. <https://doi.org/10.1007/s13204-019-01137-8>
  43. Wary RR, Baglari S, Brahma D, Gautam UK, Kalita P, et al. 2022. Synthesis, characterization, and photocatalytic activity of ZnO nanoparticles using water extract of waste coconut husk. *Environ Sci Pollut Res* 29: 42837-42848. <https://doi.org/10.1007/s11356-022-18832-9>
  44. Pérez-de-Luque A. 2020. Guest edited collection: nanotechnology in agriculture. *Sci Rep* 10: 15738. <https://doi.org/10.1038/s41598-020-73198-7>
  45. Saleem H, Zaidi SJ. 2020. Recent developments in the application of nanomaterials in agroecosystems. *Nanomaterials (Basel)* 10(12): 2411. <https://doi.org/10.3390/nano10122411>
  46. Raliya R, Saharan V, Dimkpa C, Biswas P. 2017. Nanofertilizer for precision and sustainable agriculture: current state and future perspectives. *J Agric Food Chem* 66(26): 6487-6503. <https://doi.org/10.1021/acs.jafc.7b02178>
  47. Verma ML. 2017. Enzymatic nanobiosensors in the agricultural and food industry. In: Ranjan, S., Dasgupta N, Lichtfouse E (eds) *Nanoscience in Food and Agriculture 4. Sustainable Agriculture Reviews*. Springer, pp 229-245. [https://doi.org/10.1007/978-3-319-53112-0\\_7](https://doi.org/10.1007/978-3-319-53112-0_7)
  48. Sun Y, Liang J, Tang L, Li H, Zhu Y, et al. 2019. Nano-pesticides: A great challenge for biodiversity? *Nano Today* 28: 100757. <https://doi.org/10.1016/j.nantod.2019.06.003>
  49. Prasad R, Kumar V, Prasad KS. 2014. Nanotechnology in sustainable agriculture: present concerns and future aspects. *Afr J Biotechnol* 13(6): 705-713. <https://doi.org/10.5897/AJBX2013.13554>
  50. Shah M, Fawcett D, Sharma S, Tripathy SK, Poinern GE. 2015. Green synthesis of metallic nanoparticles via biological entities. *Materials (Basel)* 8(11): 7278-7308. <https://doi.org/10.3390/ma8115377>
  51. Patil MP, Kim GD. 2018. Marine microorganisms for synthesis of metallic nanoparticles and their biomedical applications. *Colloids Surf B* 172: 487-495. <https://doi.org/10.1016/j.colsurfb.2018.09.007>
  52. Uddin AK, Siddique M, Bakar A, Rahman F, Ullah AK, et al. 2020. *Cocos nucifera* leaf extract mediated green synthesis of silver nanoparticles for enhanced antibacterial activity. *J Inorg Organomet Polym* 30(9): 3305-3316. <https://doi.org/10.1007/s10904-020-01506-9>
  53. Edmundson MC, Capeness M, Horsfall L. 2014. Exploring the potential of metallic nanoparticles within synthetic biology. *N Biotechnol* 31(6): 572-578. <https://doi.org/10.1016/j.nbt.2014.03.004>
  54. Kumar H, Bhardwaj K, Dhanjal DS, Nepovimova E, Şen F, et al. 2020. Fruit extract mediated green synthesis of metallic nanoparticles: A new avenue in pomology applications. *Int J Mol Sci* 21(22): 8458. <https://doi.org/10.3390/ijms21228458>
  55. Pantidos N, Horsfall LE. 2014. Biological synthesis of metallic nanoparticles by bacteria, fungi and plants. *J Nanomed Nanotechnol* 5: 233. <https://doi.org/10.4172/2157-7439.1000233>
  56. Jamkhande PG, Ghule NW, Bamer AH, Kalaskar MG. 2019. Metal nanoparticles synthesis: An overview on methods of preparation, advantages and disadvantages, and applications. *J Drug Deliv Sci Technol* 53: 101174. <https://doi.org/10.1016/j.jddst.2019.101174>
  57. Kato Y, Suzuki M. 2020. Synthesis of metal nanoparticles by microorganisms. *Crystals* 10(7): 589. <https://doi.org/10.3390/cryst10070589>
  58. Ali MA, Ahmed T, Wu W, Hossain A, Hafeez R, et al. 2020. Advancements in plant and microbe-based synthesis of metallic nanoparticles and their antimicrobial activity against plant pathogens. *Nanomaterials (Basel)* 10(6): 1146. <https://doi.org/10.3390/nano10061146>
  59. Usman AI, Aziz AA, Noqta OA. 2019. Application of green synthesis of gold nanoparticles: A review. *Jurnal Teknologi* 81(1): 171-182. <https://doi.org/10.11113/jt.v81.11409>
  60. Ahmed A, Usman M, Yu B, Shen Y, Cong H. 2021. Sustainable fabrication of hematite ( $\alpha$ -Fe<sub>2</sub>O<sub>3</sub>) nanoparticles using biomolecules of *Punica granatum* seed extract for unconventional solar-light-driven photocatalytic remediation of organic dyes. *Journal of Molecular Liquids* 339: 116729. <https://doi.org/10.1016/j.molliq.2021.116729>
  61. Wang W, Zhang B, Liu Q, Du P, Liu W, et al. 2018. Biosynthesis of palladium nanoparticles using *Shewanella loihica* PV-4 for excellent catalytic reduction of chromium (VI). *Environ Sci: Nano* 5(3): 730-739. <https://doi.org/10.1039/C7EN01167A>
  62. Alphandéry E. 2020. Natural metallic nanoparticles for application in nano-oncology. *Int J Mol Sci* 21(12): 4412. <https://doi.org/10.3390/ijms21124412>
  63. Lv Q, Zhang B, Xing X, Zhao Y, Cai R, et al. 2018. Biosynthesis of copper nanoparticles using *Shewanella loihica* PV-4 with antibacterial activity: Novel approach and mechanisms investigation. *J Hazard Mater* 347: 141-149. <https://doi.org/10.1016/j.jhazmat.2017.12.070>
  64. Busi S, Rajkumari J, Pattnaik S, Parasuraman P, Hnamte S. 2021. Extracellular synthesis of zinc oxide nanoparticles using *Acinetobacter schindleri* SIZ7 and its antimicrobial property against foodborne pathogens. *J Microbiol Biotech Food Sci* 5(5): 407-411. <https://doi.org/10.15414/jmbfs.2016.5.5.407-411>
  65. Rajeshkumar S, Santhoshkumar J, Vanaja M, Sivaperumal P, Ponnankikamideen M, et al. 2022. Evaluation of zebrafish toxicology and biomedical potential of *Aeromonas hydrophila* mediated copper sulfide nanoparticles. *Oxid Med Cell Longev* 2022: 7969825. <https://doi.org/10.1155/2022/7969825>
  66. Cunha FA, Cunha MD, da Frota SM, Mallmann EJ, Freire TM, et al. 2018. Biogenic synthesis of multifunctional silver nanoparticles from *Rhodotorula glutinis* and *Rhodotorula mucilaginosa*: antifungal, catalytic and cytotoxicity activities. *World J Microbiol Biotechnol* 34(9): 127. <https://doi.org/10.1007/s11274-018-2514-8>
  67. Irvani S, Varma RS. 2020. Bacteria in heavy metal remediation and nanoparticle biosynthesis. *ACS Sustainable Chem Eng* 8(14): 5395-5409. <https://doi.org/10.1021/acssuschemeng.0c00292>
  68. Iqtedar M, Riaz H, Kaleem A, Abdullah R, Aihetasham A, et al. 2020. Biosynthesis, optimization and characterization of ZnO nanoparticles using *Bacillus cereus* MN181367 and their antimicrobial activity against multidrug resistant bacteria. *Revista Mexicana De Ingenieria Química* 19(Sup. 1): 253-266. <https://doi.org/10.24275/rmiq/Bio1605>
  69. Saravanan M, Gopinath V, Chaurasia MK, Syed A, Ameen F, et al. 2018. Green synthesis of anisotropic zinc oxide nanoparticles with antibacterial and cytofriendly properties. *Microbial Pathogenesis* 115: 57-63. <https://doi.org/10.1016/j.micpath.2017.12.039>
  70. Jafari M, Rokhbakhsh-Zamin F, Shakibaie M, Moshafi MH, Ameri A, et al. 2018. Cytotoxic and antibacterial activities of biologically synthesized gold nanoparticles assisted by *Micrococcus yunnanensis* strain J2. *Biocatal Agric Biotechnol* 15: 245-253. <https://doi.org/10.1016/j.cbab.2018.06.014>
  71. Singh A, Gautam PK, Verma A, Singh V, Shivapriya PM, et al. 2020. Green synthesis of metallic nanoparticles as effective alternatives to treat antibiotics resistant bacterial infections: A review. *Biotechnology Reports* 25: e00427. <https://doi.org/10.1016/j.btre.2020.e00427>
  72. Banu AN, Balasubramanian C. 2014. Optimization and synthesis of silver nanoparticles using *Isaria fumosorosea* against human vector mosquitoes. *Parasitol Res* 113(10): 3843-3851. <https://doi.org/10.1007/s00436-014-4052-0>
  73. Taran M, Rad M, Alavi M. 2018. Biosynthesis of TiO<sub>2</sub> and ZnO nanoparticles by *Halomonas elongata* IBRC-M 10214 in different conditions of medium. *Bioimpacts* 8(2):81-89. <https://doi.org/10.15171/bi.2018.10>
  74. Monowar T, Rahman MS, Bhore SJ, Raju G, Sathasivam KV. 2018. Silver nanoparticles synthesized by using the endophytic bacterium *Pantoea ananatis* are promising antimicrobial agents against multidrug



- resistant bacteria. *Molecules* 23(12): 3220. <https://doi.org/10.3390/molecules23123220>
75. Iqtedar M, Aslam M, Akhyar M, Shehzaad A, Abdullah R, et al. 2019. Extracellular biosynthesis, characterization, optimization of silver nanoparticles (AgNPs) using *Bacillus mojavensis* BTCB15 and its antimicrobial activity against multidrug resistant pathogens. *Prep Biochem Biotechnol* 49(2): 136-142. <https://doi.org/10.1080/10826068.2018.1550654>
  76. Alsamhary KI. 2020. Eco-friendly synthesis of silver nanoparticles by *Bacillus subtilis* and their antibacterial activity. *Saudi J Biol Sci* 27(8): 2185-2191. <https://doi.org/10.1016/j.sjbs.2020.04.026>
  77. Paul D, Sinha SN. 2017. Extracellular synthesis of silver nanoparticles using *Pseudomonas aeruginosa* KUPSB12 and its antibacterial activity. *Jordan Journal of Biological Sciences* 7(4): 245-250.
  78. Gan L, Zhang S, Zhang Y, He S, Tian Y. 2018. Biosynthesis, characterization and antimicrobial activity of silver nanoparticles by a halotolerant *Bacillus endophyticus* SCU-L. *Prep Biochem Biotechnol* 48(7): 582-588. <https://doi.org/10.1080/10826068.2018.1476880>
  79. Khan R, Fulekar MH. 2016. Biosynthesis of titanium dioxide nanoparticles using *Bacillus amyloliquefaciens* culture and enhancement of its photocatalytic activity for the degradation of a sulfonated textile dye Reactive Red 31. *J Colloid Interface Sci* 475: 184-191. <https://doi.org/10.1016/j.jcis.2016.05.001>
  80. Camas M, Sazak Camas A, Kyeremeh K. 2018. Extracellular synthesis and characterization of gold nanoparticles using *Mycobacterium* sp. BRS2A-AR2 isolated from the aerial roots of the Ghanaian mangrove plant, *Rhizophora racemosa*. *Indian J Microbiol* 58(2): 214-221. <https://doi.org/10.1007/s12088-018-0710-8>
  81. Srinath BS, Namratha K, Byrappa K. 2018. Eco-friendly synthesis of gold nanoparticles by *Bacillus subtilis* and their environmental applications. *Adv Sci Lett* 24(8): 5942-5946. <https://doi.org/10.1166/asl.2018.12224>
  82. John MS, Nagoth JA, Ramasamy KP, Mancini A, Giuli G, et al. 2020. Synthesis of bioactive silver nanoparticles by a *Pseudomonas* strain associated with the antarctic psychrophilic protozoon *Euplates focardii*. *Mar Drugs* 18(1): 38. <https://doi.org/10.3390/md18010038>
  83. Ahmed E, Kalathil S, Shi L, Alharbi O, Wang P. 2018. Synthesis of ultra-small platinum, palladium and gold nanoparticles by *Sberwanella loibica* PV-4 electrochemically active biofilms and their enhanced catalytic activities. *Journal of Saudi Chemical Society* 22(8): 919-929. <https://doi.org/10.1016/j.jscs.2018.02.002>
  84. Mohamed AA, Fouda A, Abdel-Rahman MA, Hassan SE, El-Gamal MS, et al. 2019. Fungal strain impacts the shape, bioactivity and multifunctional properties of green synthesized zinc oxide nanoparticles. *Biocatalysis and Agricultural Biotechnology* 19: 101103. <https://doi.org/10.1016/j.bcab.2019.101103>
  85. Srivastava AK. 2019. The role of fungus in bioactive compound production and nanotechnology. In: Kumar A, Singh AK, Choudhary KK (eds) Role of Plant Growth Promoting Microorganisms in Sustainable Agriculture and Nanotechnology. Elsevier, Amsterdam, Netherlands pp 145-162. <https://doi.org/10.1016/B978-0-12-817004-5.00009-9>
  86. Hulikere MM, Joshi CG. 2019. Characterization, antioxidant and antimicrobial activity of silver nanoparticles synthesized using marine endophytic fungus- *Cladosporium cladosporioides*. *Process Biochemistry* 82: 199-204. <https://doi.org/10.1016/j.procbio.2019.04.011>
  87. Gudikandula K, Vadapally P, Charya MS. 2017. Biogenic synthesis of silver nanoparticles from white rot fungi: Their characterization and antibacterial studies. *OpenNano* 2: 64-78. <https://doi.org/10.1016/j.onano.2017.07.002>
  88. Abd-Elsalam KA. 2021. Fungal Nanotechnology. Special Issue: Fungal Nanotechnology. *J Fungi* 7(8): 583. <https://doi.org/10.3390/jof7080583>
  89. Zhao X, Zhou L, Riaz Rajoka MS, Yan L, Jiang C, et al. 2017. Fungal silver nanoparticles: synthesis, application and challenges. *Crit Rev Biotechnol* 38(6): 817-835. <https://doi.org/10.1080/07388551.2017.1414141>
  90. El Domany EB, Essam TM, Ahmed AE, Farghali AA. 2018. Biosynthesis physico-chemical optimization of gold nanoparticles as anti-cancer and synergetic antimicrobial activity using *Pleurotus ostrea-tus* fungus. *J Appl Pharm Sci* 8(5): 119-128. <http://doi.org/10.7324/JAPS.2018.8516>
  91. Gao Y, Arokia Vijaya Anand M, Ramachandran V, Karthikkumar V, Shalini V, et al. 2019. Biofabrication of zinc oxide nanoparticles from *Aspergillus niger*, their antioxidant, antimicrobial and anticancer activity. *J Clust Sci* 30: 937-946. <https://doi.org/10.1007/s10876-019-01551-6>
  92. Elamawi RM, Al-Harbi RE, Hendi AA. 2018. Biosynthesis and characterization of silver nanoparticles using *Trichoderma longibrachiatum* and their effect on phytopathogenic fungi. *Egypt J Biol Pest Control* 28: 28. <https://doi.org/10.1186/s41938-018-0028-1>
  93. Gupta V, Chandra N. 2020. Biosynthesis and antibacterial activity of metal oxide nanoparticles using *Brassica oleracea* subsp. botrytis (L.) leaves, an agricultural waste. *Proc Natl Acad Sci, India Sect B Biol Sci* 90: 1093-1100. <https://doi.org/10.1007/s40011-020-01184-0>
  94. AbdelRahim K, Mahmoud SY, Ali AM, Almaary KS, Mustafa AE, et al. 2017. Extracellular biosynthesis of silver nanoparticles using *Rhizopus stolonifer*. *Saudi J Biol Sci* 24(1): 208-216. <https://doi.org/10.1016/j.sjbs.2016.02.025>
  95. Xue B, He D, Gao S, Wang D, Yokoyama K, et al. 2016. Biosynthesis of silver nanoparticles by the fungus *Arthroderma fulvum* and its antifungal activity against genera of Candida, Aspergillus and Fusarium. *Int J Nanomedicine* 11: 1899-1906. <https://doi.org/10.2147/ijn.s98339>
  96. Suryavanshi P, Pandit R, Gade A, Derita M, Zachino S, et al. 2017. *Colletotrichum* sp.-mediated synthesis of sulphur and aluminium oxide nanoparticles and its in vitro activity against selected food-borne pathogens. *LWT-Food Science and Technology* 81: 188-194. <https://doi.org/10.1016/j.lwt.2017.03.038>
  97. Shamsuzzaman, Mashrai A, Khanam H, Aljawfi RN. 2017. Biological synthesis of ZnO nanoparticles using *C. albicans* and studying their catalytic performance in the synthesis of steroidal pyrazolines. *Arabian Journal of Chemistry* 10(2): S1530-S1536. <https://doi.org/10.1016/j.arabjc.2013.05.004>
  98. Fouda A, Awad MA, Eid AM, Saied E, Barghoth MG, et al. 2021. An eco-friendly approach to the control of pathogenic microbes and *Anopheles stephensi* malarial vector using magnesium oxide nanoparticles (Mg-nps) fabricated by *Penicillium chrysogenum*. *Int J Mol Sci* 22(10): 5096. <https://doi.org/10.3390/ijms22105096>
  99. Fathy RM, Mahfouz AY. 2021. Eco-friendly graphene oxide-based magnesium oxide nanocomposite synthesis using fungal fermented by-products and gamma rays for outstanding antimicrobial, antioxidant, and anticancer activities. *J Nanostruct Chem* 11(2): 301-321. <https://doi.org/10.1007/s40097-020-00369-3>
  100. Naimi-Shamel N, Pourali P, Dolatabadi S. 2019. Green synthesis of gold nanoparticles using *Fusarium oxysporum* and antibacterial activity of its tetracycline conjugant. *J Mycol Med* 29(1): 7-13. <https://doi.org/10.1016/j.mycmed.2019.01.005>
  101. Kobashigawa JM, Robles CA, Ricci ML, Carmarán CC. 2018. Influence of strong bases on the synthesis of silver nanoparticles (AgNPs) using the ligninolytic fungi *Trametes trogii*. *Saudi J Biol Sci* 26(7): 1331-1337. <https://doi.org/10.1016/j.sjbs.2018.09.006>
  102. Abdel-Hadi AM, Awad MF, Abo-Dahab NF, Elkady MF. 2014. Extracellular synthesis of silver nanoparticles by *Aspergillus terreus*: biosynthesis, characterization and biological activity. *Biosci Biotechnol Res Asia* 11(3): 1179-1186. <http://doi.org/10.13005/bbra/1503>
  103. Al-Zubaidi S, Al-Ayafi A, Abdelkader H. 2019. Biosynthesis, characterization and antifungal activity of silver nanoparticles by *Aspergillus niger* isolate. *Journal of Nanotechnology Research* 2(2019): 22-35.
  104. Devi LS, Joshi SR. 2015. Ultrastructures of silver nanoparticles biosynthesized using endophytic fungi. *J Microsc Ultrastruct* 3(1): 29-37. <https://doi.org/10.1016/j.jmau.2014.10.004>
  105. Gupta K, Chundawat TS, Malek NA. 2020. Antibacterial, antifungal, photocatalytic activities and seed germination effect of mycosynthesized silver nanoparticles using *Fusarium oxysporum*. *Biointerface Res Appl Chem* 11(4): 12082-12091. <https://doi.org/10.33263/BRI-AC114.1208212091>

106. Guilger-Casagrande M, Germano-Costa T, Pasquoto-Stigliani T, Fraceto LF, Lima RD. 2019. Biosynthesis of silver nanoparticles employing *Trichoderma harzianum* with enzymatic stimulation for the control of *Sclerotinia sclerotiorum*. *Sci Rep* 9: 14351. <https://doi.org/10.1038/s41598-019-50871-0>
107. Mishra A, Kumari M, Pandey S, Chaudhry V, Gupta KC, et al. 2014. Biocatalytic and antimicrobial activities of gold nanoparticles synthesized by *Trichoderma* sp. *Bioresour Technol* 166: 235-242. <https://doi.org/10.1016/j.biortech.2014.04.085>
108. Bin-Jumah M, Monera AA, Albasher G, Alarifi S. 2020. Effects of green silver nanoparticles on apoptosis and oxidative stress in normal and cancerous human hepatic cells in vitro. *Int J Nanomedicine* 15: 1537-1548. <https://doi.org/10.2147/ijn.s239861>
109. Apriandanu DO, Yulizar Y. 2019. *Tinospora crispa* leaves extract for the simple preparation method of CuO nanoparticles and its characterization. *Nano-Structures & Nano-Objects* 20: 100401. <https://doi.org/10.1016/j.nanoso.2019.100401>
110. Jayapriya M, Dhanasekaran D, Arulmozhi M, Nandhakumar E, Senthilkumar N, et al. 2019. Green synthesis of silver nanoparticles using *Piper longum* catkin extract irradiated by sunlight: antibacterial and catalytic activity. *Res Chem Intermed* 45(6): 3617-3631. <https://doi.org/10.1007/s11164-019-03812-5>
111. Kumar TS. 2013. Physical and chemical characterization of biomaterials. In: Bandyopadhyay A, Bose S (eds) Characterization of biomaterials. Academic Press, pp 11-47. <https://doi.org/10.1016/B978-0-12-415800-9.00002-4>
112. Nayak S, Sajankila SP, Rao CV, Hegde AR, Mutalik S. 2021. Biogenic synthesis of silver nanoparticles using *Jatropha curcas* seed cake extract and characterization: evaluation of its antibacterial activity. *Energy sources, Part A: Recovery, Utilization, And Environmental Effects* 43(24): 1-9. <https://doi.org/10.1080/15567036.2019.1632394>
113. Vijilvani C, Bindhu MR, Frincy FC, AlSalhi MS, Sabitha S, et al. 2020. Antimicrobial and catalytic activities of biosynthesized gold, silver and palladium nanoparticles from *Solanum nigrum* leaves. *J Photochem Photobiol B Biol* 202: 111713. <https://doi.org/10.1016/j.jphotobiol.2019.111713>
114. Ajitha B, Reddy YA, Jeon HJ, Ahn CW. 2018. Synthesis of silver nanoparticles in an eco-friendly way using *Phyllanthus amarus* leaf extract: antimicrobial and catalytic activity. *Adv Powder Technol* 29(1): 86-93. <https://doi.org/10.1016/j.apt.2017.10.015>
115. Francis S, Nair KM, Paul N, Koshy EP, Mathew B. 2019. Catalytic activities of green synthesized silver and gold nanoparticles. *Materials Today: Proceedings* 9(1): 97-104. <https://doi.org/10.1016/j.matpr.2019.02.042>
116. Korani S, Rashidi K, Hamelian M, Jalalvand AR, Tajehmiri A, et al. 2021. Evaluation of antimicrobial and wound healing effects of gold nanoparticles containing *Abelmoschus esculentus* (L.) aqueous extract. *Bioinorg Chem* 2021: 7019130. <https://doi.org/10.1155/2021/7019130>
117. Syed A, Ahmad A. 2012. Extracellular biosynthesis of platinum nanoparticles using the fungus *Fusarium oxysporum*. *Colloids Surf B Biointerfaces* 97: 27-31. <https://doi.org/10.1016/j.colsurfb.2012.03.026>
118. Nguyen VT, Bowyer MC, Van Altna IA, Scarlett CJ. 2017. Microwave-assisted extraction as an advanced technique for optimization of saponin yield and antioxidant potential from *Phyllanthus amarus*. *Separation Science and Technology* 52(17): 1-11. <https://doi.org/10.1080/01496395.2017.1374972>
119. Zahra F, Utami FA, Girsang GC, Mulya SZ, Fentiana VD, et al. 2020. Economic evaluation of zinc oxide nanoparticle production through green synthesis method using *Cassia fistula* plant extract. *IJECA* 5(2): 18-24. <https://doi.org/10.47238/ijeca.v5i2.133>
120. Mittal AK, Chisti Y, Banerjee UC. 2013. Synthesis of metallic nanoparticles using plant extracts. *Biotechnology Advances* 31(2): 346-356. <https://doi.org/10.1016/j.biotechadv.2013.01.003>
121. Salayová A, Bedlovičová Z, Daneu N, Baláz M, Lukáčová Bujňáková Z, et al. 2021. Green synthesis of silver nanoparticles with antibacterial activity using various medicinal plant extracts: Morphology and antibacterial efficacy. *Nanomaterials (Basel)* 11(4): 1005. <https://doi.org/10.3390/nano11041005>
122. Jadoun S, Arif R, Jangid NK, Meena RK. 2021. Green synthesis of nanoparticles using plant extracts: A review. *Environ Chem Lett* 19(1): 355-374. <https://doi.org/10.1007/s10311-020-01074-x>
123. Lee SH, Salunke BK, Kim BS. 2014. Sucrose density gradient centrifugation separation of gold and silver nanoparticles synthesized using *Magnolia kobus* plant leaf extracts. *Biotechnol Bioproc E* 19: 169-174. <https://doi.org/10.1007/s12257-013-0561-4>
124. Shankar S, Jaiswal L, Aparna RS, Prasad RG. 2014. Synthesis, characterization, in vitro biocompatibility, and antimicrobial activity of gold, silver and gold silver alloy nanoparticles prepared from *Lansium domesticum* fruit peel extract. *Materials Letters* 137: 75-78. <https://doi.org/10.1016/j.matlet.2014.08.122>
125. Siddiqi KS, Husen A. 2016. Green synthesis, characterization and uses of palladium/platinum nanoparticles. *Nanoscale Res Lett* 11(1): 482. <https://doi.org/10.1186/s11671-016-1695-z>
126. Akintelu SA, Folorunso AS, Folorunso FA, Oyebamiji AK. 2020. Green synthesis of copper oxide nanoparticles for biomedical application and environmental remediation. *Heliyon* 6(7): e04508. <https://doi.org/10.1016/j.heliyon.2020.e04508>
127. Sharma D, Kumar N, Mehrotra T, Pervaiz N, Agrawal L, et al. 2021. In vitro and in silico molecular docking studies of Rheum emodi-derived diamagnetic SnO<sub>2</sub> nanoparticles and their cytotoxic effects against breast cancer. *New J Chem* 45(3): 1695-1711. <https://doi.org/10.1039/D0NJ04670A>
128. Sharma D, Parveen K, Oza A, Ledwani L. 2018. Synthesis of anthraquinone-capped TiO<sub>2</sub> nanoparticles using *R. emodi* roots: preparation, characterization and cytotoxic potential. *Rend Fis Acc Lincei* 29: 649-658. <https://doi.org/10.1007/s12210-018-0696-5>
129. Goutam SP, Saxena G, Singh V, Yadav AK, Bharagava RN, et al. 2018. Green synthesis of TiO<sub>2</sub> nanoparticles using leaf extract of *Jatropha curcas* L. for photocatalytic degradation of tannery wastewater. *Chem Eng J* 336: 386-396. <https://doi.org/10.1016/j.cej.2017.12.029>
130. Parveen K, Kumar N, Ledwani L. 2022. Green synthesis of Zinc Oxide nanoparticles mediated from *Cassia renigera* bark and detect its effects on four varieties of rice. *ChemistrySelect* 7(17): e202200415. <https://doi.org/10.1002/slct.202200415>
131. Abd El-Aziz AR, Al-Othman MR. 2019. Gold nanoparticles biosynthesis using *zingiber officinale* and their impact on the growth and chemical composition of lentil (*Lens culinaris* medic.). *Pak J Bot* 51(2): 443-450. [https://doi.org/10.30848/PJB2019-2\(21\)](https://doi.org/10.30848/PJB2019-2(21))
132. Letsholathebe D, Thema FT, Mphale K, Maabong K, Magdalane CM. 2021. Green synthesis of ZnO doped *Moringa oleifera* leaf extract using Tiron yellow dye for photocatalytic applications. *Materials Today: Proceedings* 36(Part 2): 475-479. <https://doi.org/10.1016/j.matpr.2020.05.119>
133. Sukri SN, Shameli K, Wong MM, Teow SY, Chew J, et al. 2019. Cytotoxicity and antibacterial activities of plant-mediated synthesized zinc oxide (ZnO) nanoparticles using *Punica granatum* (pomegranate) fruit peels extract. *J Mol Struct* 1189: 57-65. <https://doi.org/10.1016/j.molstruc.2019.04.026>
134. Nguyen TM, Nguyen TA, Tuong-Van Pham N, Ly QV, Tran TT, et al. 2021. Biosynthesis of metallic nanoparticles from waste *Passiflora edulis* peels for their antibacterial effect and catalytic activity. *Arab J Chem* 14(4): 103096. <https://doi.org/10.1016/j.arabjc.2021.103096>
135. Brindhadevi K, Samuel MS, Verma TN, Vasantharaj S, Sathiyavimal S, et al. 2020. Zinc oxide nanoparticles (ZnONPs) -induced antioxidants and photocatalytic degradation activity from hybrid grape pulp extract (HGPE). *Biocatal Agric Biotechnol* 28: 101730. <https://doi.org/10.1016/j.cbac.2020.101730>
136. Srivastava V, Sharma YC, Sillanpää M. 2015. Green synthesis of magnesium oxide nanoflower and its application for the removal of divalent metallic species from synthetic wastewater. *Ceramics International* 41(5)



- Part B): 6702-6709. <https://doi.org/10.1016/j.ceramint.2015.01.112>
137. Krishnaswamy K, Vali H, Orsat V. 2014. Value-adding to grape waste: Green synthesis of gold nanoparticles. *J Food Eng* 142: 210-220. <https://doi.org/10.1016/j.jfoodeng.2014.06.014>
  138. Zamani A, Marjani AP, Mousavi Z. 2019. Agricultural waste bio-mass-assisted nanostructures: Synthesis and application. *Green Process Synth* 8(1): 421-429. <https://doi.org/10.1515/gps-2019-0010>
  139. Sangeetha J, Thangadurai D, Hospet R, Purushotham P, Manowade KR, et al. 2017. Production of bionanomaterials from agricultural wastes. In: Prasad R, Kumar M, Kumar V (eds) *Nanotechnology*. Springer, Singapore, pp 33-58. [https://doi.org/10.1007/978-981-10-4573-8\\_3](https://doi.org/10.1007/978-981-10-4573-8_3)
  140. Omran BA, Aboelazayem O, Nassar HN, El-Salamony RA, El-Gendy NS. 2021. Biovalorization of mandarin waste peels into silver nanoparticles and activated carbon. *Int J Environ Sci Technol* 18: 1119-1134. <https://doi.org/10.1007/s13762-020-02873-z>
  141. Ibrahim HMM. 2015. Green synthesis and characterization of silver nanoparticles using banana peel extract and their antimicrobial activity against representative microorganisms. *JRRAS* 8(3): 265-275. <https://doi.org/10.1016/j.jrras.2015.01.007>
  142. Kumar R, Roopan SM, Prabhakarn A, Khanna VG, Chakroborty S. 2012. Agricultural waste *Annona squamosa* peel extract: biosynthesis of silver nanoparticles. *Spectrochim Acta A Mol Biomol Spectrosc* 90: 173-176. <https://doi.org/10.1016/j.saa.2012.01.029>
  143. Dungani R, Abdul Khalil HPS, Aprilia NAS, Sumardi I, Aditiawati P, et al. 2017. Bionanomaterial from agricultural waste and its application. In: Jawaid M, Boufi S, Abdul Khalil HPS (eds) *Cellulose-reinforced nanofibre composites*. Woodhead Publishing, pp 45-88. <https://doi.org/10.1016/B978-0-08-100957-4.00003-6>
  144. Yang N, Li WH, Hao L. 2014. Biosynthesis of Au nanoparticles using agricultural waste mango peel extract and its in vitro cytotoxic effect on two normal cells. *Materials Letters* 134: 67-70. <https://doi.org/10.1016/j.matlet.2014.07.025>
  145. Thi TU, Nguyen TT, Thi YD, Thi KH, Phan BT, et al. 2020. Green synthesis of ZnO nanoparticles using orange fruit peel extract for antibacterial activities. *RSC Adv* 10(40): 23899-23907. <https://doi.org/10.1039/D0RA04926C>
  146. Doan VD, Phung MT, Nguyen TL, Mai TC, Nguyen TD. 2020. Noble metallic nanoparticles from waste *Nypa fruticans* fruit husk: biosynthesis, characterization, antibacterial activity and recyclable catalysis. *Arab J Chem* 13(10): 7490-7503. <https://doi.org/10.1016/j.arabjc.2020.08.024>
  147. Lakshminpathy R, Palakshi Reddy B, Sarada NC, Chidambaram K, Khadeer Pasha SK. 2015. Watermelon rind-mediated green synthesis of noble palladium nanoparticles: catalytic application. *Appl Nanosci* 5: 223-228. <https://doi.org/10.1007/s13204-014-0309-2>
  148. Debnath G, Das P, Saha AK. 2019. Green synthesis of silver nanoparticles using mushroom extract of *Pleurotus giganteus*: characterization, antimicrobial, and  $\alpha$ -amylase inhibitory activity. *BioNanoSci* 9: 611-619. <https://doi.org/10.1007/s12668-019-00650-y>
  149. Fatimah IS. 2018. Biosynthesis and characterization of ZnO nanoparticles using rice bran extract as low-cost templating agent. *J Eng Sci Technol* 13(2): 409-420.
  150. Roopan SM, Madhumitha G, Rahuman AA, Kamaraj C, Bharathi A, et al. 2013. Low-cost and eco-friendly phyto-synthesis of silver nanoparticles using *Cocos nucifera* coir extract and its larvicidal activity. *Ind Crops Prod* 43: 631-635. <https://doi.org/10.1016/j.indcrop.2012.08.013>
  151. Athinarayanan J, Periasamy VS, Alhazmi M, Alataiah KA, Alshatwi AA. 2015. Synthesis of biogenic silica nanoparticles from rice husks for biomedical applications. *Ceramics International* 41(1): 275-281. <https://doi.org/10.1016/j.ceramint.2014.08.069>
  152. Sinsinwar S, Sarkar MK, Suriya KR, Nithyanand P, Vadivel V. 2018. Use of agricultural waste (coconut shell) for the synthesis of silver nanoparticles and evaluation of their antibacterial activity against selected human pathogens. *Microb Pathog* 124: 30-37. <https://doi.org/10.1016/j.micpath.2018.08.025>
  153. Khanna P, Kaur A, Goyal D. 2019. Algae-based metallic nanoparticles: Synthesis, characterization and applications. *J Microbiol Methods* 163: 105656. <https://doi.org/10.1016/j.jmimet.2019.105656>
  154. Makuła P, Pacia M, Macyk W. 2018. How to correctly determine the band gap energy of modified semiconductor photocatalysts based on UV-Vis spectra. *J Phys Chem Lett* 9(23): 6814-6817. <https://doi.org/10.1021/acs.jpcllett.8b02892>
  155. Wang N, Sun Q, Yu J. 2019. Ultrasmall metal nanoparticles confined within crystalline nanoporous materials: a fascinating class of nanocatalysts. *Advanced Materials* 31(1): 1803966. <https://doi.org/10.1002/adma.201803966>
  156. Shnoudeh AJ, Hamad I, Abdo RW, Qadumii L, Jaber AY, et al. 2019. Synthesis, characterization, and applications of metal nanoparticles. In: Tekade RK (ed) *Biomaterials and Bionanotechnology*. Academic Press, pp 527-612. <https://doi.org/10.1016/B978-0-12-814427-5.00015-9>
  157. Mourdikoudis S, Pallares RM, Thanh NT. 2018. Characterization techniques for nanoparticles: comparison and complementarity upon studying nanoparticle properties. *Nanoscale* 10(27): 12871-12934. <https://doi.org/10.1039/C8NR02278J>
  158. Titus D, Samuel EJJ, Roopan SM. 2019. Nanoparticle characterization techniques. In: Shukla AK, Iravani S (eds) *Green synthesis, characterization and applications of nanoparticles*. Elsevier, pp 303-319. <https://doi.org/10.1016/B978-0-08-102579-6.00012-5>
  159. Gao X, Lowry GV. 2018. Progress towards standardized and validated characterizations for measuring physicochemical properties of manufactured nanomaterials relevant to nano health and safety risks. *Nano-Impact* 9: 14-30. <https://doi.org/10.1016/j.impact.2017.09.002>
  160. Callahan PG, Stinville JC, Yao ER, Echlin MP, Titus MS, et al. 2018. Transmission scanning electron microscopy: Defect observations and image simulations. *Ultramicroscopy* 186: 49-61. <https://doi.org/10.1016/j.ultramicro.2017.11.004>
  161. Larue C, Castillo-Michel H, Stein RJ, Fayard B, Pouyet E, et al. 2016. Innovative combination of spectroscopic techniques to reveal nanoparticle fate in a crop plant. *Spectrochim Acta B: At Spectrosc* 119: 17-24. <https://doi.org/10.1016/j.sab.2016.03.005>
  162. Muñoz-Fernandez L, Alkan G, Milošević O, Rabanal ME, Friedrich B. 2017. Synthesis and characterisation of spherical core-shell Ag/ZnO nanocomposites using single and two - steps ultrasonic spray pyrolysis (USP). *Catalysis Today* 321-322: 26-33. <https://doi.org/10.1016/j.cattod.2017.11.029>
  163. Holder CF, Schaak RE. 2019. Tutorial on powder X-ray diffraction for characterizing nanoscale materials. *ACS Nano* 13(7): 7359-7365. <https://doi.org/10.1021/acsnano.9b05157>
  164. dos Santos J, de Oliveira RS, de Oliveira TV, Velho MC, Konrad MV, et al. 2021. 3D printing and nanotechnology: a multiscale alliance in personalized medicine. *Adv Funct Mater* 31(16): 2009691. <https://doi.org/10.1002/adfm.202009691>
  165. Xiang Y, Zheng G, Liang Z, Jin Y, Liu X, et al. 2020. Visualizing the growth process of sodium microstructures in sodium batteries by in-situ  $^{23}\text{Na}$  MRI and NMR spectroscopy. *Nat Nanotechnol* 15: 883-890. <https://doi.org/10.1038/s41565-020-0749-7>
  166. Adams FC, Barbante C. 2013. Nanoscience, nanotechnology and spectrometry. *Spectrochim Acta B: At Spectrosc* 86: 3-13. <https://doi.org/10.1016/j.sab.2013.04.008>
  167. Modena MM, Rühle B, Burg TP, Wuttke S. 2019. Nanoparticle characterization: what to measure? *Advanced Materials* 31(32): 1901556. <https://doi.org/10.1002/adma.201901556>
  168. Begum R, Farooqi ZH, Naseem K, Ali F, Batool M, et al. 2018. Applications of UV/Vis spectroscopy in characterization and catalytic activity of noble metal nanoparticles fabricated in responsive polymer microgels: a review. *Crit Rev Anal Chem* 48(6): 503-516. <https://doi.org/10.1080/10408347.2018.1451299>
  169. Sundar S, Venkatachalam G, Kwon SJ. 2018. Biosynthesis of copper oxide (CuO) nanowires and their use for the electrochemical sensing of dopamine. *Nanomaterials (Basel)* 8(10): 823. <https://doi.org/10.3390/nano8100823>



170. Gemmi M, Mugnaioli E, Gorelik TE, Kolb U, Palatinus L, et al. 2019. 3D electron diffraction: the nanocrystallography revolution. *ACS Cent Sci* 5(8): 1315-1329. <https://doi.org/10.1021/acscentsci.9b00394>
171. Ditta A. 2012. How helpful is nanotechnology in agriculture? *Adv Nat Sci: Nanosci Nanotechnol* 3: 033002. <http://doi.org/10.1088/2043-6262/3/3/033002>
172. Prasad R, Kumar V, Prasad KS. 2014. Nanotechnology in sustainable agriculture: present concerns and future aspects. *Afr J Biotechnol* 13(6): 705-713. <https://doi.org/10.5897/AJBX2013.13554>
173. Xu L, Shoaie N, Jahanpeyma F, Zhao J, Azimzadeh M, et al. 2020. Optical, electrochemical and electrical (nano) biosensors for detection of exosomes: A comprehensive overview. *Biosens Bioelectron* 161: 112222. <https://doi.org/10.1016/j.bios.2020.112222>
174. Christopher FC, Kumar PS, Christopher FJ, Joshiba GJ, Madhesh P. 2020. Recent advancements in rapid analysis of pesticides using nano biosensors: a present and future perspective. *Journal of Cleaner Production* 269: 122356. <https://doi.org/10.1016/j.jclepro.2020.122356>
175. Solaimuthu A, Vijayan AN, Murali P, Korrapati PS. 2020. Nano-biosensors and their relevance in tissue engineering. *Curr Opin Biomed Eng* 13: 84-93. <https://doi.org/10.1016/j.cobme.2019.12.005>
176. Ghaffar N, Farrukh MA, Naz S. 2020. Applications of nanobiosensors in agriculture. In: Javad S (ed) *Nanoagronomy*. Springer, Cham, pp 179-196. [https://doi.org/10.1007/978-3-030-41275-3\\_10](https://doi.org/10.1007/978-3-030-41275-3_10)
177. Elahi N, Kamali M, Baghersad MH, Amini B. 2019. A fluorescence Nano-biosensors immobilization on Iron (MNPs) and gold (AuNPs) nanoparticles for detection of *Shigella spp*. *Mater Sci Eng C* 105: 110113. <https://doi.org/10.1016/j.msec.2019.110113>
178. Huang X, Zhu Y, Kianfar E. 2021. Nano biosensors: properties, applications and electrochemical techniques. *J Mater Res Technol* 12: 1649-1672. <https://doi.org/10.1016/j.jmrt.2021.03.048>
179. Kaushal M, Wani SP. 2017. Nanosensors: frontiers in precision agriculture. In: Prasad R, Kumar M, Kumar V (eds) *Nanotechnology*. Springer, Singapore, pp 279-291. [https://doi.org/10.1007/978-981-10-4573-8\\_13](https://doi.org/10.1007/978-981-10-4573-8_13)
180. Singh RP. 2017. Application of nanomaterials toward development of nanobiosensors and their utility in agriculture. In: Prasad R, Kumar M, Kumar V (eds) *Nanotechnology*. Springer, Singapore, pp 293-303. [https://doi.org/10.1007/978-981-10-4573-8\\_14](https://doi.org/10.1007/978-981-10-4573-8_14)
181. Panpatte DG, Jhala YK. 2019. *Nanotechnology for agriculture: advances for sustainable agriculture*. Springer Nature, Singapore. <https://doi.org/10.1007/978-981-32-9370-0>
182. Shawon ZB, Hoque ME, Chowdhury SR. 2020. Nanosensors and nanobiosensors: agricultural and food technology aspects. In: Pal K, Gomes F (eds) *Micro and nano technologies, nanofabrication for smart nanosensor application*, Elsevier, pp 135-161. <https://doi.org/10.1016/B978-0-12-820702-4.00006-4>
183. Hussein HS, Shaarawy HH, Hussien NH, Hawash SI. 2019. Preparation of nano-fertilizer blend from banana peels. *Bull Natl Res Cent* 43: 26. <https://doi.org/10.1186/s42269-019-0058-1>
184. León-Silva S, Arrieta-Cortes R, Fernández-Luqueño F, López-Valdez F. 2018. Design and production of nanofertilizers. In: López-Valdez F, Fernández-Luqueño F (eds) *Agricultural nanobiotechnology*. Springer, Cham, pp 17-31. [https://doi.org/10.1007/978-3-319-96719-6\\_2](https://doi.org/10.1007/978-3-319-96719-6_2)
185. Subramanian KS, Thirunavukkarasu M. 2017. Nano-fertilizers and nutrient transformations in soil. In: Ghorbanpour M, Manika K, Varma A (eds) *Nanoscience and plant-soil systems*. Springer, Cham, pp 305-319. [https://doi.org/10.1007/978-3-319-46835-8\\_11](https://doi.org/10.1007/978-3-319-46835-8_11)
186. Zulfiqar F, Navarro M, Ashraf M, Akram NA, Munné-Bosch S. 2019. Nanofertilizer use for sustainable agriculture: Advantages and limitations. *Plant Sci* 110270. <https://doi.org/10.1016/j.plantsci.2019.110270>
187. Lateef A, Nazir R, Jamil N, Alam S, Shah R, et al. 2019. Synthesis and characterization of environmentally friendly corn cob biochar based nano-composite - A potential slow release nano-fertilizer for sustainable agriculture. *Environ Nanotechnol Monit Manag* 100212. <https://doi.org/10.1016/j.enmm.2019.100212>
188. Yadi M, Mostafavi E, Saleh B, Davaran S, Aliyeva I, et al. 2018. Current developments in green synthesis of metallic nanoparticles using plant extracts: a review. *Artif Cells Nanomed Biotechnol* 46(sup 3): S336-S343. <https://doi.org/10.1080/21691401.2018.1492931>
189. Shebl A, Hassan AA, Salama DM, Abd El-Aziz ME, AbdElwahed MS. 2019. Green synthesis of Nanofertilizers and their application as a foliar for *Cucurbita pepo* L.. *J Nanomater* 2019: 3476347. <https://doi.org/10.1155/2019/3476347>
190. Kah M. 2015. Nanopesticides and nanofertilizers: emerging contaminants or opportunities for risk mitigation?. *Front Chem* 3: 64. <https://doi.org/10.3389/fchem.2015.00064>
191. Basavegowda N, Baek KH. 2021. Current and future perspectives on the use of nanofertilizers for sustainable agriculture: the case of phosphorus nanofertilizer. *3 Biotech* 11(7): 357. <https://doi.org/10.1007/s13205-021-02907-4>
192. Al-Mamun MR, Hasan MR, Ahommed MS, Bacchu MS, Ali MR, et al. 2021. Nanofertilizers towards sustainable agriculture and environment. *Environ Technol Innov* 23: 101658. <https://doi.org/10.1016/j.eti.2021.101658>
193. Fatima F, Hashim A, Anees S. 2020. Efficacy of nanoparticles as nanofertilizer production: a review. *Environ Sci Pollut Res Int* 28(2): 1292-1303. <https://doi.org/10.1007/s11356-020-11218-9>
194. Agathokleous NP. 2020. Nano-pesticides: a great challenge for biodiversity? The need for a broader perspective. *Nano Today* 30: 100808. <https://doi.org/10.1016/j.nantod.2019.100808>
195. Priyanka P, Kumar D, Yadav K, Yadav A. 2019. Nanopesticides: synthesis, formulation and application in agriculture. In: Abd-ElSalam K, Prasad R (eds) *Nanobiotechnology applications in plant protection*. Springer, Cham, pp 129-143. [https://doi.org/10.1007/978-3-030-13296-5\\_7](https://doi.org/10.1007/978-3-030-13296-5_7)
196. Gahukar RT, Das RK. 2020. Plant-derived nanopesticides for agricultural pest control: challenges and prospects. *Nanotechnol Environ Eng* 5(1): 3. <https://doi.org/10.1007/s41204-020-0066-2>
197. Grillo R, Fraceto LF, Amorim MJ, Scott-Fordsmand JJ, Schoonjans R, et al. 2020. Ecotoxicological and regulatory aspects of environmental sustainability of nanopesticides. *J Hazard Mater* 404: 124148. <https://doi.org/10.1016/j.jhazmat.2020.124148>
198. Bhagyaraj SM, Oluwafemi OS. 2018. Nanotechnology: the science of the invisible. In: Bhagyaraj SM, Oluwafemi OS, Kalarikkal N, Thomas S (eds) *Micro and nano technologies, synthesis of inorganic nanomaterials*. Woodhead Publishing, Sawston, pp 1-18. <https://doi.org/10.1016/B978-0-08-101975-7.00001-4>
199. Li L, Xu Z, Kah M, Lin D, Filser J. 2019. Nanopesticides: a comprehensive assessment of environmental risk is needed before widespread agricultural application. *Environ Sci Technol* 53(14): 7923-7924. <https://doi.org/10.1021/acs.est.9b03146>
200. Boddula R, Trivedi U, Pothu R, Rajput MS, Saran A. 2019. Nanopesticides and nanosensors in agriculture. In: Prasad R (ed) *Nanotechnology in the life sciences*. Springer, Cham, pp 165-181. [https://doi.org/10.1007/978-3-030-12496-0\\_8](https://doi.org/10.1007/978-3-030-12496-0_8)
201. Fraceto LF, De Castro VL, Grillo R, Ávila D, Oliveira HC, et al. 2020. *Nanopesticides*. Springer Nature, Switzerland. <https://doi.org/10.1007/978-3-030-44873-8>
202. Baudhdh K, Kumar S, Singh RP, Korstad J. 2020. Ecological and practical applications for sustainable agriculture. Springer Nature, Switzerland. <https://doi.org/10.1007/978-981-15-3372-3>
203. Shalaby TA, Bayoumi Y, Abdalla N, Taha H, Alshaal T, et al. 2016. Nanoparticles, soils, plants and sustainable agriculture. In: Ranjan, S., Dasgupta N, Lichtfouse E (eds) *Nanoscience in food and agriculture 1. sustainable agriculture reviews*. Springer, Cham, pp 283-312. [https://doi.org/10.1007/978-3-319-39303-2\\_10](https://doi.org/10.1007/978-3-319-39303-2_10)