

# Plant-Extract Mediated Synthesis of Copper- and Iron-Based Nanoparticles for Various Applications

Harshita Sachdeva<sup>1</sup>, Naveen Sharma<sup>1</sup>, Amit Sharma<sup>1</sup>, and Rahul Shrivastava<sup>2</sup>

<sup>1</sup>Department of Chemistry, University of Rajasthan, Jaipur, Rajasthan, India

<sup>2</sup>Department of Chemistry, Manipal University Jaipur, Rajasthan, India

## \*Correspondence to:

Harshita Sachdeva

Department of Chemistry, University of Rajasthan, Jaipur, Rajasthan, India

E-mail: [drhmsachdevaster@gmail.com](mailto:drhmsachdevaster@gmail.com)

Rahul Shrivastava

Department of Chemistry, Manipal University Jaipur, Rajasthan, India

E-mail: [chem.rahul@gmail.com](mailto:chem.rahul@gmail.com)

Received: August 23, 2022

Accepted: November 04, 2022

Published: November 06, 2022

**Citation:** Sachdeva H, Sharma N, Sharma A, Shrivastava R. 2022. Plant-Extract Mediated Synthesis of Copper-and Iron-Based Nanoparticles for Various Applications. *NanoWorld J* 8(S1): S162-S167.

**Copyright:** © 2022. Sachdeva et al. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC-BY) (<http://creativecommons.org/licenses/by/4.0/>) which permits commercial use, including reproduction, adaptation, and distribution of the article provided the original author and source are credited.

Published by United Scientific Group

## Abstract

Green synthesis has attracted wide attention as a trustworthy and environmentally friendly approach for synthesising a broad variety of nanomaterials, including metal nanoparticles NPs and metal oxide NPs. It is considered an advanced tool for reducing the harsh effects related to the conventional methods of NPs synthesis, which are generally used in the laboratory and industry. The current review article focuses on investigations indicating the new processes to produce copper (Cu) and iron (Fe)-based NPs by using extracts of plant parts, particularly, flower, stem, seeds, whole plant, and fruit extracts, followed by the future opportunity of work in this area.

## Keywords

Green synthesis, Nanomaterials, Plant extracts, Copper, Iron-based

## Introduction

In recent years, nanotechnology has appeared as a modern and revolutionary technology with applicability in a wide range of fields [1-6]. Amongst nanomaterials, current research focus is mainly on the synthesis of NPs since they can be easily produced and manipulated. Magnetic NPs come under the category of inorganic NPs, which can be applied in different areas due to their numerous exceptional characteristics. Since the last decade, new approaches for the synthesis of nanomaterials have been an exciting area of exploration in nanoscience and nanotechnology [7-10]. Several chemical and physical methods are reported in the literature for NPs preparation [11-13] but these methods suffer from various drawbacks, hence research attention has recently moved in the direction of developing clean and eco-pleasant synthetic approaches [14-16]. Keeping in view of the disadvantages related to physical and chemical methods of NPs synthesis (toxicity, recyclability, stability requirements, etc.), green synthetic approaches have been invented, which may utilize plant extracts, bacteria, fungi, algae, etc. Among the available green methodologies for NPs synthesis, employment of extracts of different plant parts is quite simple for large scale production in comparison with bacteria or fungi-mediated synthesis. This may be due to the valuable phytochemicals present in various natural plant extracts, which have the ability to reduce metal salts into metal NPs. There are excellent review articles published since last 2-3 years regarding the preparation of Fe and Cu-based NPs by applying green methods [17-19]. In continuation, we would like to report the current findings and progress in this particular area. As a result of the limitations associated with chemical and physical methods of NPs synthesis, we decided to provide a brief overview of recent developments rather than a comprehensive review. In this review article, green synthetic approaches

for Fe and Cu-based NPs using natural extracts of plants have been summarized.

## Green Production of Copper-Based Nanoparticles

NPs possess a broad application in different areas of recent research significance, including environmental, pharmaceutical, biomedical, energy, and catalytic applications, etc. [20-22]. Among transition-metal based NPs, copper nanoparticles (CuNPs) are of great concern because they are cheap and easily available [23, 24]. They can be utilized in heat transfer systems, sensors, as catalysts for chemical reactions, and as antimicrobial agents [25-32]. There are many reported methods for metal NPs synthesis [33, 34] which cause production of pure and well-defined NPs, yet they are neither profitable nor environmentally pleasant due to the toxic nature of chemicals. Hence, biosynthesis of nanoparticles is one of the important tools of green chemistry, which makes use of a diverse range of plants, fungus, bacteria, yeast, and viruses [35, 36].

It is well-known that different plant parts can be used for the preparation of Cu-based NPs using different precursor salts under varying reaction conditions. In this regard, Nasrollahzadeh et al. [37] caused immobilization of CuNPs on a natrolite zeolite surface by using *Anthemis xylopoda* flower aqueous extract, which acts as a reducing and stabilising agent. The performance of the catalyst was examined for N-formylation of amines under eco-friendly reaction conditions. It was observed that the catalyst could be reused at least five times without significant loss of activity. Further, *Punica granatum* seeds extract arbitrated CuNPs were prepared by Nazar et al. [38]. The CuNPs displayed excellent photocatalytic activity for the methylene blue (MB) degradation under irradiation of solar light. Another study reported by Ismail [39] revealed inexpensive and ecological method for CuNPs production using fruit extract of sumaq (*Rhus coriaria* L.) as a stabilizing agent. CuNPs showed excellent activity against Gram-positive bacteria. It was established that *Citrofortunella microcarpa* leaves extract synthesized CuO NPs were employed as photocatalyst for the purification of rhodamine B dye polluted water [40]. Further, *Ailanthus altissima* leaf aqueous extract prepared CuO NPs demonstrated significant inhibitory activity against *Staphylococcus aureus* and *Escherichia coli* [41]. *Jatropha curcas* leaf extract utilized green synthesis of CuNPs was described by Ghosh et al. [42]. It was illustrated that the flavonoids, glycosides, tannins, and alkaloids present in the plant extract were used as reducing, stabilizing, and capping agents. CuNPs synthesized by using *Hagenia abyssinica* (Brace) JF. Gmel leaf extract appear to be more crystalline in nature as reported by Ananda Murthy et al. [43]. Asemanni M and Anarjan N [44] synthesized CuO NPs (mean particle size of 80 nm) using walnut leaf extract. Another study by Sinha et al. [45] described the biogenic synthesis of spherical CuNPs (22 - 27 nm) by using the peel extracts of *Citrus grandis*. As revealed by X-ray diffraction (XRD) data, the average crystallite size was 18 nm. Further, Mohamed et al. [46] reported a simple, beneficial, and environment-friendly method using seedless dates'

extract rich in phenolics and flavonoids for synthesizing Cu/CuO NPs. Furthermore, Akhter et al. [47] prepared CuNPs from stem extract of holoparasitic plants (*Orobancha aegyptiaca*) which were found to be active against *E. coli* and *S. aureus* and exhibit nematicidal activity. Another report by Sukumar et al. [48] showed that *Caesalpinia bonducella* seed extract synthesized CuO NPs could be used as a graphite electrode surface modifier and also possesses high antibacterial activity. Sarker et al. [49] prepared spherical, mono-disperse, and highly stable CuO NPs in a unique and economical way by using *Adiantum lunulatum* whole plant extract.

Another literature report by Rabiee et al. [50] demonstrated CuO NPs synthesis by using leaf extracts of *Achillea millefolium*. Investigation of the catalytic activity of CuO NPs for azide-alkyne cycloaddition and A<sup>3</sup> coupling reaction, and also the photocatalytic activity was examined for the degradation of MB dye. Ethanolic and aqueous extracts of hawthorn fruit were chosen to prepare monodispersed and stable CuNPs by Długosz et al. [51]. Recently, Nieto-Maldonado et al. [52] synthesized CuNPs using extracts of fresh petals of *Rosa* 'double delight' or fresh leaves of *Gardenia jasminoides* Ellis as stabilizing agents. A simple, clean, and efficient process for synthesizing CuNPs was further developed by Shubhashree et al. [53] by using *Hyptis suaveolens* (L.) extracts. Furthermore, Tamil Elakkiya et al. [54] synthesized CuO NPs using *Sesbania aculeate* leaf extract, which increased the growth of *Brassica nigra* and exhibited powerful antimicrobial activity. Table 1 indicates synthesis of Cu-based NPs using various plant extracts.

## Green Production of Iron-Based Nanoparticles

Iron oxide nanoparticles (IONPs) synthesis is in progress due to their unique properties including super-paramagnetism, biocompatibility, non-toxic nature, as well as biodegradability, cost-effective production, etc. [55, 56]. Investigations into the Fe-based NPs preparation via plant extracts have been thoroughly examined by many authors [57-60].

FeNPs are commonly synthesized using sodium borohydride (NaBH<sub>4</sub>), which is quite expensive and not eco-compatible. Hence, different tea extracts were used for FeNPs synthesis via the green route [61, 62]. The polyphenols present in tea catalysed the synthesis of Fe NPs. Furthermore, IONPs functionalized bio-nanocomposite prepared by using pomegranate leaf extract [63] was found to be efficient in removing hexavalent chromium. Additional application of FeNPs synthesised from *Eucalyptus* leaf extracts includes its use in wastewater treatment [64]. It is well known that fruit peels discarded in the production of fruit juice cause environmental pollution. These peels can be utilized for the production of FeNPs for various applications. In this regard, *Citrus maxima* peel extracts synthesized FeNPs are used for the reduction of Fe (III) in aqueous solution [65]. Further, *Hibiscus sabdariffa* Roselle flower synthesized FeNPs catalysed the oxidative degradation of rhodamine B [66]. A simple and fast procedure for the synthesis of spherical Fe<sub>2</sub>O<sub>3</sub> NPs using fruit extracts of Cornelian cherry was reported by Rostamizadeh et al. [67]. These Fe<sub>2</sub>O<sub>3</sub> NPs are found to be more effective in stimulating

**Table 1:** Synthesis of copper-based nanoparticles using plant extracts.

S. No.	Plant	Reducing or capping agent	Size (nm)	Structure /Shape	References
1.	<i>Punica granatum</i>	Seed extract	43.9	Semi-spherical	[38]
2.	<i>Rhuscoriaria L.</i>	Fruit extract	22	Semi-spherical	[39]
3.	<i>Citrofortunella macrocarpa</i>	Leaf extract	54 - 68	Spherical	[40]
4.	<i>Citrus grandis</i>	Peel extract	22 - 27	Spherical	[45]
5.	<i>Ailanthus altissima</i>	Leaf extract	20	Spherical	[41]
6.	<i>Hagenia abyssinica</i>	Leaf extract	34.76	Mix. of spherical hexagonal, triangular cylindrical and irregular	[43]
7.	<i>Adiantum lunulatum</i>	Plant extract	1.5 - 20	Spherical	[49]
8.	<i>Achillea millefolium</i>	Leaf extracts	28	Semi-spherical	[50]
9.	<i>Hawthorn berries</i>	Fruit extract	200	Spherical	[51]
10.	<i>Rosa 'double delight'</i>	Fresh petals	11.7	Spherical	[52]
11.	<i>Gardenia jasminoides</i> Ellis	Fresh leaves	3	Spherical	[52]

the growth of barley than its bulk. Another catalytic application of the magnetic NPs synthesized by Prasad et al. using watermelon rinds includes the preparation of bioactive tetrahydro-pyrimidine derivatives [68]. This is eco-friendly green method for the synthesis of tetrahydro-pyrimidine derivatives in high yield (94%) with only 5 mmol of  $Fe_3O_4$  NPs as catalyst without the requirement of additional reductant or surfactant. Furthermore, Yew et al. reported the preparation of magnetite NPs by using seaweed, *Kappaphycus alvarezii* [69]. In one more study, Ardakani et al. [70] synthesized spherical FeNPs using aqueous leaf extract of *Chlorophytum comosum*, which can be utilized to control the removal of methyl orange from the aqueous solution.

Nanoscience plays a very important role in controlling air pollution and in the degradation of dyes. In this perspective, Karpagavinayagam and Vedhi [71] developed a non-toxic and clean method for the synthesis of IONPs using *Avicennia marina* flower extract. Table 2 indicates synthesis of various iron-based NPs using natural plant extracts.

## Future Perspectives

This review highlighted current developments in the green synthetic protocols using plant extracts for Fe and Cu-based NPs and their modifications with respect to particular application in catalysis ranging from organic transformations to electrocatalysis, photocatalysis, and so on. Cu-based NPs synthesized by using various plant extracts exhibit excellent photocatalytic activity for the degradation of methylene blue and for the purification of rhodamine B dye polluted water and CuNPs were found to be active against *S. aureus* and *E. coli*. They also exhibit nematicidal activity and could be used as a graphite electrode surface modifier. Fe-based NPs are efficient in removing hexavalent chromium and can be used for wastewater treatment. Further, they can be used to catalyse the oxidative degradation of rhodamine B, removal of methyl orange from the aqueous solution, preparation of bioactive tetrahydro-pyrimidine derivatives and to control air pollution. These investigations suggest that Fe and Cu-based nanomaterials are promising candidates for environmental, biomedical, and catalytic applications. Further, more studies are required regarding the synthesis and functionalization

**Table 2:** Synthesis of iron-based nanoparticles using plant extracts.

S. No.	Plant	Reducing or capping agent	Size (nm)	Structure /Shape	References
1.	<i>Hibiscus sabdariffa</i>	Flower extract	18	Spherical	[66]
2.	<i>Citrullus lanatus</i>	Rind extract	2 - 20	Spherical	[68]
3.	<i>Kappaphycus alvarezii</i>	Seaweed extract	14.7	Spherical	[69]
4.	<i>Chlorophytum comosum</i>	Leaf extract	Below 100	Spherical	[70]
5.	<i>Avicennia marina</i>	Flower extract	10 - 40	Honeycomb	[71]



of Cu and Fe-based nanoparticles for particular application.

## Conflict of Interest

Authors declare that they have no conflict of interest.

## Acknowledgements

For providing the department with the required research facilities, the authors thank the head of the chemistry department at the University of Rajasthan in Jaipur and Manipal University Jaipur, Rajasthan, India.

## References

- Singh J, Dutta T, Kim KH, Rawat M, Samddar P, et al. 2018. Green synthesis of metals and their oxide nanoparticles: applications for environmental remediation. *J Nanobiotechnology* 16(1): 84. <https://doi.org/10.1186/s12951-018-0408-4>
- Smith DM, Simon JK, Baker Jr. JR. 2013. Applications of nanotechnology for immunology. *Nat Rev Immunol* 13(8): 592-605. <https://doi.org/10.1038/nri3488>
- Banin U, Waiskopf N, Hammarström L, Boschloo G, Freitag M, et al. 2020. Nanotechnology for catalysis and solar energy conversion. *Nanotechnology* 32(4): 42003. <https://doi.org/10.1088/1361-6528/abbce8>
- Davoodi S, Al-Shargabi M, Wood DA, Rukavishnikov VS, Minaev KM. 2022. Experimental and field applications of nanotechnology for enhanced oil recovery purposes: a review. *Fuel* 324: 124669. <https://doi.org/10.1016/j.fuel.2022.124669>
- Nasrollahzadeh M, Sajadi SM, Sajjadi M, Issaabadi Z. 2019. Applications of nanotechnology in daily life. In: Nasrollahzadeh M, Sajadi SM, Sajjadi M, Issaabadi Z, Atarod M (eds) *Interface Science and Technology*. Elsevier, pp 28: 113-43. <https://doi.org/10.1016/B978-0-12-813586-0.00004-3>
- Anjum S, Ishaque S, Fatima H, Farooq W, Hano C, et al. 2021. Emerging applications of nanotechnology in healthcare systems: grand challenges and perspectives. *Pharmaceuticals (Basel)* 14(8): 707. <https://doi.org/10.3390/ph14080707>
- O'Neal DP, Hirsch LR, Halas NJ, Payne JD, West JL. 2004. Photo-thermal tumor ablation in mice using near infrared-absorbing nanoparticles. *Cancer Lett* 209: 171-176. <https://doi.org/10.1016/j.canlet.2004.02.004>
- Gour A, Jain NK. 2019. Advances in green synthesis of nanoparticles. *Artif Cells Nanomed Biotechnol* 47(1): 844-851. <https://doi.org/10.1080/21691401.2019.1577878>
- Meng X, Fan K, Yan X. 2019. Nanozymes: an emerging field bridging nanotechnology and enzymology. *Sci China Life Sci* 62(11): 1543-1546. <https://doi.org/10.1007/s11427-019-1557-8>
- Salem SS, Fouda A. 2021. Green synthesis of metallic nanoparticles and their prospective biotechnological applications: an overview. *Biol Trace Elem Res* 199(1): 344-370. <https://doi.org/10.1007/s12011-020-02138-3>
- Jadoun S, Arif R, Jangid NK, Meena RK. 2021. Green synthesis of nanoparticles using plant extracts: a review. *Environ Chem Lett* 19(1): 355-374. <https://doi.org/10.1007/s10311-020-01074-x>
- Jamkhande PG, Ghule NW, Bamer AH, Kalaskar MG. 2019. Metal nanoparticles synthesis: An overview on methods of preparation, advantages and disadvantages, and applications. *J Drug Deliv Sci Technol* 53: 101174. <https://doi.org/10.1016/j.jddst.2019.101174>
- Baig N, Kammakakam I, Falath W. 2021. Nanomaterials: a review of synthesis methods, properties, recent progress, and challenges. *Mater Adv* 2(6): 1821-1871. <https://doi.org/10.1039/D0MA00807A>
- Shah M, Fawcett D, Sharma S, Tripathy SK, Poinern GE. 2015. Green synthesis of metallic nanoparticles via biological entities. *Materials Basel* 8(11): 7278-308. <https://doi.org/10.3390/ma8115377>
- Chandra H, Kumari P, Bontempi E, Yadav S. 2020. Medicinal plants: treasure trove for green synthesis of metallic nanoparticles and their biomedical applications. *Biocatal Agric Biotechnol* 24: 101518. <https://doi.org/10.1016/j.bcab.2020.101518>
- Adelere IA, Lateef A. 2016. A novel approach to the green synthesis of metallic nanoparticles: the use of agro-wastes, enzymes, and pigments. *Nanotechnol Rev* 5(6): 567-87. <https://doi.org/10.1515/ntrev-2016-0024>
- Yadwade R, Kirtiwar S, Ankamwar B. 2021. A review on green synthesis and applications of iron oxide nanoparticles. *J Nanosci Nanotechnol* 21(12): 5812-5834. <https://doi.org/10.1166/jnn.2021.19285>
- Waris A, Din M, Ali A, Ali M, Afridi S, et al. 2021. A comprehensive review of green synthesis of copper oxide nanoparticles and their diverse biomedical applications. *Inorg Chem Commun* 123: 108369. <https://doi.org/10.1016/j.inoche.2020.108369>
- Bouafia A, Laouini SE, Ouahrani MR. 2020. A review on green synthesis of CuO nanoparticles using plant extract and evaluation of antimicrobial activity. *Asian Journal of Research in Chemistry* 13(1): 65-70. <https://doi.org/10.5958/0974-4150.2020.00014.0>
- De M, Ghosh PS, Rotello VM. 2008. Applications of nanoparticles in biology. *Adv Mater* 20(22): 4225-4241. <https://doi.org/10.1002/adma.200703183>
- Stark WJ, Stoessel PR, Wohlleben W, Hafner AJ. 2015. Industrial applications of nanoparticles. *Chem Soc Rev* 44(16): 5793-805. <https://doi.org/10.1039/C4CS00362D>
- Han X, Xu K, Taratula O, Farsad K. 2019. Applications of nanoparticles in biomedical imaging. *Nanoscale* 11(3): 799-819. <https://doi.org/10.1039/c8nr07769j>
- Dang TMD, Le TTT, Fribourg-Blanc E, Dang MC. 2012. Influence of surfactant on the preparation of silver nanoparticles by polyol method. *Adv Nat Sci Nanosci Nanotechnol* 3(3): 035004. <https://doi.org/10.1088/2043-6262/3/3/035004>
- Din MI, Rehan R. 2017. Synthesis, characterization, and applications of copper nanoparticles. *Anal Lett* 50(1): 50-62. <https://doi.org/10.1080/00032719.2016.1172081>
- Wang Z, Zhu L, Sun S, Wang J, Yan W. 2021. One-dimensional nanomaterials in resistive gas sensor: from material design to application. *Chemosensors* 9(8): 198. <https://doi.org/10.3390/chemosensors9080198>
- Steinhauer S. 2021. Gas sensors based on copper oxide nanomaterials: a review. *Chemosensors* 9(3): 51. <https://doi.org/10.3390/chemosensors9030051>
- Okonkwo EC, Wole-Osho I, Almanassra IW, Abdullatif YM, Al-Ansari T. 2021. An updated review of nanofluids in various heat transfer devices. *J Therm Anal Calorim* 145(6): 2817-2872. <https://doi.org/10.1007/s10973-020-09760-2>
- Guo Y, Wen M, Li G, An T. 2021. Recent advances in VOC elimination by catalytic oxidation technology onto various nanoparticles catalysts: a critical review. *Appl Catal B* 281: 119447. <https://doi.org/10.1016/j.apcatb.2020.119447>
- Pasinszki T, Krebsz M. 2020. Synthesis and application of zero-valent iron nanoparticles in water treatment, environmental remediation, catalysis, and their biological effects. *Nanomaterials (Basel)* 10(5): 917. <https://doi.org/10.3390/nano10050917>
- Iravani S, Varma RS. 2020. Sustainable synthesis of cobalt and cobalt oxide nanoparticles and their catalytic and biomedical applications. *Green Chem* 22(9): 2643-2661. <https://doi.org/10.1039/D0GC00885K>
- Wang X, Dong Q, Qiao H, Huang Z, Saray MT, et al. 2020. Continuous synthesis of hollow high entropy nanoparticles for energy and catalysis applications. *Adv Mater* 32(46): 2002853. <https://doi.org/10.1002/adma.202002853>
- Magaye R, Zhao J, Bowman L, Ding M. 2012. Genotoxicity and carcinogenicity of cobalt-, nickel- and copper-based nanoparticles. *Exp*

- Ther Med* 4(4): 551-561. <https://doi.org/10.3892/etm.2012.656>
33. Rajput N. 2015. Methods of preparation of nanoparticles-a review. *Int J Adv Eng Technol* 7(6): 1806.
34. Iravani S, Korbekandi H, Mirmohammadi SV, Zolfaghari B. 2014. Synthesis of silver nanoparticles: chemical, physical and biological methods. *Res Pharm Sci* 9(6): 385-406.
35. Hussain I, Singh NB, Singh A, Singh H, Singh SC. 2016. Green synthesis of nanoparticles and its potential application. *Biotechnol Lett* 38(4): 545-60. <https://doi.org/10.3892/etm.2012.656>
36. Belwal S, Saritha R, Sachdeva H, Kiran G. 2019. Synthesis, characterization and prediction of anticancer potentiality of some novel green nanoparticles by molecular docking and ADMET techniques. *Bull Chem Soc Ethiop* 33(3): 493-504. <https://doi.org/10.4314/bcse.v33i3.10>
37. Nasrollahzadeh M, Sajadi SM, Hatamifard AJ. 2015. *Anthemisis xylopada* flowers aqueous extract assisted in situ green synthesis of Cu nanoparticles supported on natural Natrolite zeolite for N-formylation of amines at room temperature under environmentally benign reaction conditions. *J Colloid Interface Sci* 460: 146-153. <https://doi.org/10.1016/j.jcis.2015.08.040>
38. Nazar N, Bibi I, Kamal S, Iqbal M, Nouren S, et al. 2018. Cu nanoparticles synthesis using biological molecule of *P granatum* seeds extract as reducing and capping agent: Growth mechanism and photo-catalytic activity. *Int J Biol Macromol* 106: 1203-1210. <https://doi.org/10.1016/j.ijbiomac.2017.08.126>
39. Ismail MI. 2020. Green synthesis and characterizations of copper nanoparticles. *Mater Chem Phys* 240: 122283. <https://doi.org/10.1016/j.matchemphys.2019.122283>
40. Rafique M, Shafiq F, Gillani SS, Shakil M, Tahir MB, et al. 2020. Eco-friendly green and biosynthesis of copper oxide nanoparticles using *Citrofortunella microcarpa* leaves extract for efficient photocatalytic degradation of Rhodamin B dye form textile wastewater. *Optik* 208: 164053. <https://doi.org/10.1016/j.ijleo.2019.164053>
41. Awwad A, Amer M, Al-aqarbeh M. 2020. TiO<sub>2</sub>-kaolinite nanocomposite prepared from the Jordanian Kaolin clay: Adsorption and thermodynamics of Pb (II) and Cd (II) ions in aqueous solution. *Chemistry International* 6(4): 168-148. <https://doi.org/10.5281/zenodo.3597558>
42. Ghosh MK, Sahu S, Gupta I, Ghorai TK. 2020. Green synthesis of copper nanoparticles from an extract of *Jatropha curcas* leaves: characterization, optical properties, CT-DNA binding and photocatalytic activity. *RSC Adv* 10(37): 22027- 22035. <https://doi.org/10.1039/D0RA03186K>
43. Murthy HCA, Desalegn T, Kassa M, Abebe B, Assefa T. 2020. Synthesis of green copper nanoparticles using medicinal plant *Hagenia abyssinica* (Brace) JF. Gmel. leaf extract: Antimicrobial properties. *J Nanomater* 2020: 3924081. <https://doi.org/10.1155/2020/3924081>
44. Asemanni M, Anarjan N. 2019. Green synthesis of copper oxide nanoparticles using *Juglans regia* leaf extract and assessment of their physico-chemical and biological properties. *Green Processing and Synthesis* 8(1): 557-567. <https://doi.org/10.1515/gps-2019-0025>
45. Sinha T, Ahmaruzzaman M. 2015. Biogenic synthesis of Cu nanoparticles and its degradation behavior for methyl red. *Mater Lett* 159: 168-171. <https://doi.org/10.1016/j.matlet.2015.06.099>
46. Mohamed EA. 2020. Green synthesis of copper & copper oxide nanoparticles using the extract of seedless dates. *Heliyon* 6(1): e03123. <https://doi.org/10.1016/j.heliyon.2019.e03123>
47. Akhter G, Khan A, Ali SG, Khan TA, Siddiqi KS, et al. 2020. Antibacterial and nematocidal properties of biosynthesized Cu nanoparticles using extract of holoparasitic plant. *SN Appl Sci* 2(7): 1268. <https://doi.org/10.1007/s42452-020-3068-6>
48. Sukumar S, Rudrasenan A, Nambiar DP. 2020. Green-synthesized rice-shaped copper oxide nanoparticles using *Caesalpinia bonducella* seed extract and their applications. *ACS Omega* 5(2): 1040-1051. <https://doi.org/10.1021/acsomega.9b02857>
49. Sarkar J, Chakraborty N, Chatterjee A, Bhattacharjee A, Dasgupta D, et al. 2020. Green synthesized copper oxide nanoparticles ameliorate defence and antioxidant enzymes in *Lens culinaris*. *Nanomaterials (Basel)* 10(2): 312. <https://doi.org/10.3390/nano10020312>
50. Rabiee N, Bagherzadeh M, Kiani M, Ghadiri AM, Etesamifard F, et al. 2020. Biosynthesis of copper oxide nanoparticles with potential biomedical applications. *Int J Nanomedicine* 15: 3983-3999. <https://doi.org/10.2147/IJN.S255398>
51. Długosz O, Chwastowski J, Banach M. 2020. Hawthorn berries extract for the green synthesis of copper and silver nanoparticles. *Chem Pap* 74(1): 239-252. <https://doi.org/10.1007/s11696-019-00873-z>
52. Nieto-Maldonado A, Bustos-Guadarrama S, Espinoza-Gomez H, Flores-López LZ, Ramirez-Acosta K, et al. 2022. Green synthesis of copper nanoparticles using different plant extracts and their antibacterial activity. *J Environ Chem Eng* 10(2): 107130. <https://doi.org/10.1016/j.jece.2022.107130>
53. Shubhashree KR, Reddy R, Gangula AK, Nagananda GS, Badiya PK, et al. 2022. Green synthesis of copper nanoparticles using aqueous extracts from *Hyptis suaveolens* (L.). *Mater Chem Phys* 280: 125795.
54. Elakkiya VT, Meenakshi RV, Kumar PS, Karthik V, Shankar KR, et al. 2022. Green synthesis of copper nanoparticles using *Sesbania aculeata* to enhance the plant growth and antimicrobial activities. *Int J Environ Sci Technol* 19(3): 1313-1322. <https://doi.org/10.1007/s13762-021-03182-9>
55. Latha N, Gowri M. 2014. Bio synthesis and characterisation of Fe<sub>3</sub>O<sub>4</sub> nanoparticles using *Caricaya papaya* leaves extract. *International Journal of Science and Research* 3(11): 1551-1556.
56. Lu X, Zhou H, Liang Z, Feng J, Lu Y, et al. 2022. Biodegradable and biocompatible exceedingly small magnetic iron oxide nanoparticles for T1-weighted magnetic resonance imaging of tumors. *J Nanobiotechnol* 20(1): 350. <https://doi.org/10.1186/s12951-022-01562-y>
57. Bouafia A, Laouini SE. 2021. Plant-mediated synthesis of iron oxide nanoparticles and evaluation of the antimicrobial activity: a review. *Mini Rev Org Chem* 18(6): 725-734. <https://doi.org/10.2174/1570193X17999200908091139>
58. Jacinto MJ, Silva VC, Valladão DM, Souto RS. 2021. Biosynthesis of magnetic iron oxide nanoparticles: a review. *Biotechnol Lett* 43(1): 1-2. <https://doi.org/10.1007/s10529-020-03047-0>
59. Hamdy NM, Boseila AA, Ramadan A, Basalious EB. 2022. Iron oxide nanoparticles-plant insignia synthesis with favorable biomedical activities and less toxicity, in the “Era of the-Green”: a systematic review. *Pharmaceutics* 14(4): 844. <https://doi.org/10.3390/pharmaceutics14040844>
60. Fahmy HM, Mohamed FM, Marzouq MH, Mustafa AB, Alsoudi AM, et al. 2018. Review of green methods of iron nanoparticles synthesis and applications. *BioNanoSci* 8(2): 491-503. <https://doi.org/10.1007/s12668-018-0516-5>
61. Huang L, Weng X, Chen Z, Megharaj M, Naidu R. 2014. Green synthesis of iron nanoparticles by various tea extracts: comparative study of the reactivity. *Spectrochim Acta A Mol Biomol Spectrosc* 130: 295-301. <https://doi.org/10.1016/j.saa.2014.04.037>
62. Hoag GE, Collins JB, Holcomb JL, Hoag JR, Nadagouda MN, et al. 2009. Degradation of bromothymol blue by ‘greener’ nano-scale zero-valent iron synthesized using tea polyphenols. *J Mater Chem* 19(45): 8671-8677. <https://doi.org/10.1039/B909148C>
63. Rao A, Bankar A, Kumar AR. 2013. Removal of hexavalent chromium ions by *Yarrowia lipolytica* cells modified with phyto-inspired Fe0 nanoparticles. *J Contam Hydrol* 146: 63-73. <https://doi.org/10.1016/j.jconhyd.2012.12.008>
64. Wang T, Jin X, Chen Z, Megharaj M, Naidu R. 2014. Green synthesis of Fe nanoparticles using eucalyptus leaf extracts for treatment of

- eutrophic wastewater. *Sci Total Environ* 466: 210-213. <https://doi.org/10.1016/j.scitotenv.2013.07.022>
65. Wei Y, Fang Z, Zheng L, Tan L, Tsang EP. 2016. Green synthesis of Fe nanoparticles using *Citrus maxima* peels aqueous extracts. *Mater Lett* 185: 384-386. <https://doi.org/10.1016/j.matlet.2016.09.029>
66. Khan Z, Al-Thabaiti SA. 2018. Green synthesis of zero-valent Fe-nanoparticles: catalytic degradation of rhodamine B, interactions with bovine serum albumin and their enhanced antimicrobial activities. *J Photochem Photobiol B* 180: 259-267. <https://doi.org/10.1016/j.jphotobiol.2018.02.017>
67. Rostamizadeh E, Iranbakhsh A, Majd A. 2020. Green synthesis of Fe<sub>2</sub>O<sub>3</sub> nanoparticles using fruit extract of *Cornus mas* L. and its growth-promoting roles in Barley. *J Nanostruct Chem* 10: 125-130. <https://doi.org/10.1007/s40097-020-00335-z>
68. Prasad C, Gangadhara S, Venkateswarlu P. 2016. Bio-inspired green synthesis of Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles using watermelon rinds and their catalytic activity. *Appl Nanosci* 6(6): 797-802. <https://doi.org/10.1007/s13204-015-0485-8>
69. Yew YP, Shameli K, Miyake M, Kuwano N, Khairudin NB, et al. 2016. Green synthesis of magnetite (Fe<sub>3</sub>O<sub>4</sub>) nanoparticles using seaweed (*Kappaphycus alvarezii*) extract. *Nanoscale Res Lett* 11(1): 276. <https://doi.org/10.1186/s11671-016-1498-2>
70. Ardakani LS, Alimardani V, Tamaddon AM, Amani AM, Taghizadeh S. 2021. Green synthesis of iron-based nanoparticles using *Chlorophytum comosum* leaf extract: methyl orange dye degradation and antimicrobial properties. *Heliyon* 7(2): e06159. <https://doi.org/10.1016/j.heliyon.2021.e06159>
71. Karpagavinayagam P, Vedhi C. 2019. Green synthesis of iron oxide nanoparticles using *Avicennia marina* flower extract. *Vacuum* 160: 286-292. <https://doi.org/10.1016/j.vacuum.2018.11.043>